Acute Lymphoblastic Leukemia in Adults

Effective Date: October 2023
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Background

Acute Lymphoblastic Leukemia (ALL) is a highly aggressive hematological malignancy resulting from the proliferation and expansion of lymphoid blasts in the blood, bone marrow and other organs.\(^1\) ALL occurs with a bimodal distribution with an early peak in children 4 – 5 years old followed by a second peak at ~ 50 years of age\(^2\) with the worldwide incidence being ~ 1 – 4.75/100,000 individuals with a male:female prevalence of roughly 1:3:1.\(^1\) It is the most common childhood acute leukemia accounting for ~ 80% of the pediatric leukemias but contributing to only 20% of adult leukemias. Although significant progress has been made in treating adult ALL the overall survival amongst adults 18 to 60 years old is only 35% in contrast to childhood ALL in which overall survival at five years is more than 80%.\(^1\)

Over the past two decades the treatment of adult ALL has changed significantly with the introduction of pediatric protocols for the treatment of adolescents and young adults, the addition of tyrosine kinase (TKI) inhibitors for the treatment of Philadelphia positive/BCR-ABL positive ALL, a reevaluation of the role of allogeneic stem cell transplantation for standard risk ALL patients, the incorporation of minimal residual disease into risk assessments and most recently the introduction of novel agents such blinatumomab, inotuzumab and chimeric antigen receptor – T cell therapy for relapsed/refractory ALL.

Guideline Questions

1. How are adult patients with ALL diagnosed and worked-up?
2. What are the classifications and prognostications of adult ALL?
3. What are the recommended treatment approaches to ALL?

Search Strategy

The PubMed database was searched for relevant studies, guidelines and consensus documents published using the search term 'Acute Lymphoblastic Leukemia'. Clinical trials, clinical practice guidelines, systematic reviews and meta analyses written in English were included.

Target Population

The following recommendations apply to adult cancer patients with suspicion or diagnosis of ALL.

Summary of Recommendations

Diagnosis and Work-up

1. All patients suspected of leukemia should undergo bone marrow studies incorporating morphological assessment, immunophenotyping, cytogenetic +/- FISH and molecular evaluation.
   a. For B-cell ALL, results of BCR-ABL by PCR or t(9;22) by cytogenetics/FISH should be available within 5 days as this will influence the induction treatment regimen used.
b. Patients with failed cytogenetics for B-cell ALL should have molecular/FISH testing for BCR-ABL (if not yet done) and MLL (KMT2A) rearrangement.

c. A diagnostic baseline lumbar puncture should be performed with cell count and morphologic evaluation of the CSF cytospin preparation. If the result is unclear, flow cytometry should be performed.

2. If bone marrow studies are not feasible peripheral blood should be sent for immunophenotyping, cytogenetics and molecular studies.

3. Patients for whom anthracycline based treatment is contemplated should receive a cardiac evaluation e.g. MUGA scan or echocardiogram or cardiac MRI.

4. Transplant – eligible patients and their siblings should be HLA typed.

**Classification and Prognostication**

1. Patients should be classified as having B-cell or T-cell ALL based upon immunophenotyping results.

2. Pre-treatment risk stratification should be ascertained for all patients using age and cytogenetics/FISH and/or molecular studies.

3. Post-treatment risk stratification should include the outcomes of minimal residual disease assessment using either flow cytometry or PCR (see below)

**Principles of Treatment Initiation**

1. Induction therapy should only be administered in a leukemia centre with physician and nursing expertise in the management of acute leukemias.

2. Induction therapy should not be initiated until the BCR-ABL status is known. If the patient is symptomatic, a steroid pre-phase should be indicated.

3. Initiation of induction therapy should be accompanied by tumour lysis syndrome (TLS) prophylaxis, including hydration, urate lowering agents and close monitoring of TLS chemistries.

**Treatment of Ph/BCR-ABL negative ALL**

1. Eligible adults under age 60 should be treated with a pediatric-based protocol.
   a. In Alberta the current standard regimen is the modified Dana Farber Cancer Institute (DFCI) protocol.
   b. Patients with co-morbidities, or those unable to tolerate the full DFCI protocol, may be treated with a less intensive regimen, such as the modified DFCI protocol for patients age 60 or over.

2. Fit adults age 60-75 should be treated with curative intent.
   a. The Princess Margaret Hospital modified DFCI protocol for adults above age 60 has produced favourable results in this population compared to other regimens and may be used. Other curative-intent regimens are also acceptable.

3. Patients over age 75, or those under age 75 with major co-morbidities precluding intensive chemotherapy, should be considered for palliative chemotherapy with corticosteroids and
vincristine +/- low-dose asparaginase, followed by low-dose maintenance chemotherapy if CR is achieved.

**Treatment of Ph/BCR-ABL positive ALL**

1. Ph/BCR-ABL positive patients who are fit for chemotherapy should be treated with a BCR-ABL tyrosine kinase inhibitor (TKI) combined with induction and post-remission therapy.
   a. A less intensive induction regimen with corticosteroids, vincristine and TKI (e.g. Chalandon protocol) is preferred for initial induction, as it produces higher CR rates due to lower induction mortality. In younger, fit patients, this should be followed by intensification (e.g. HyperCVAD Part B + TKI, as per the Chalandon protocol).
   b. Patients achieving a hematologic CR should be continued on post-remission chemotherapy + TKI. This may consist of the modified Princess Margaret Hospital DFCI CNS, intensification and maintenance phases, or HyperCVAD + TKI. Asparaginase should not be used due to increased toxicity when used concurrently with TKI.
   c. Imatinib (600 – 800mg per day), is currently the standard first line TKI drug.
      i. Patients with intolerance to, or not achieving an adequate response to, imatinib should be switched to a second generation TKI such as dasatinib
      ii. Ponatinib should be used in patients with a T315I mutation
      iii. For patients with CNS disease at diagnosis, dasatinib should be used upfront due to its superior CNS penetration.
   d. Patients with Ph/BCR-ABL positive ALL who are elderly, or otherwise unfit for intensive chemotherapy or transplant, should be treated with the Chalandon induction protocol cycle A, or corticosteroids + TKI, followed by low-dose maintenance chemotherapy + TKI.
   e. TKIs should be continued indefinitely in patients who are not transplanted.
   f. Patients not transplanted should be closely monitored for disease progression with serial PCR testing every 3 months. Reappearance of PCR positivity, if confirmed, should prompt a change in TKI and referral for allogeneic HSCT, if a potential candidate.
   g. Patients with persistence of, or reappearance of BCR-ABL transcripts by PCR, should have mutational testing, specifically to look for the presence of a T315I mutation.

**Role of MRD (minimal or measurable residual disease) assessments**

1. Ph/BCR-ABL negative patients should have MRD assessments following induction chemotherapy, or by week 16, by flow cytometry or molecular techniques.
   a. MRD positive B-ALL patients, defined as > 0.1% of mononuclear cells by flow (>10^{-3}), should receive immuno-therapy using 1-2 cycles of blinatumomab with an intent to achieve MRD negativity.
   b. MRD positive T-ALL patients should receive intensified chemotherapy with an intent to achieve MRD negativity.
c. MRD positive patients post-intensive induction or at week 16, defined as > 0.1% of mononuclear cells by flow, should be considered for allogeneic HSCT in CR1, as these patients are at higher risk of relapse.

2. Ph/BCR-ABL positive patients should have MRD assessments following induction chemotherapy, or by week 16, by quantitative PCR.
   a. Patients who are MRD positive by PCR at the end of a two-cycle induction using the Chalandon protocol, or by week 16 using other protocols, should be switched to a second generation TKI such as dasatinib. If already on a 2nd generation TKI, the patient should be switched to ponatinib. The TKI should be combined with either chemotherapy or blinatumomab.
   b. MRD positive patients at these timepoints should be referred for allogeneic HSCT in CR1, as per below.

Role of allogeneic stem cell transplantation
1. Allogeneic HSCT should not be routinely performed in patients with Ph/BCR-ABL negative ALL in CR-1. However, patients with the following high-risk features should be considered for HSCT:
   i. t(4;11) with MLL (KMT2A) rearrangement
   ii. Early T-cell precursor (ETP) ALL
   iii. Lack of attainment of hematologic CR with first induction
   iv. MRD positivity at end of induction or by week 16, as per above.
   v. Inability to deliver sufficient asparaginase dosing during intensification therapy
   a. There is no clear evidence that other cytogenetic abnormalities, or specific cell surface markers, constitute high risk features when using a pediatric-based regimen
   b. It is also not clear whether patients with a high-risk feature at baseline who achieve MRD negativity after induction therapy require a transplant.

2. Fit BCR-ABL positive patients up to age 70 may be considered for allogeneic HSCT in CR-1.
   a. Patients with BCR-ABL positivity by PCR, either post-induction or by week 16 (depending on the regimen), should be referred for allogeneic HSCT, as per 6.1.3. If an HLA matched related or unrelated donor is unavailable, a haploidentical transplant should be considered.
   b. Patients achieving early MRD negativity by PCR, either post-induction or by week 16 (depending on the regimen), may be continued on post-induction chemotherapy together with TKI without a transplant. Transplant is also an option in these patients.
   c. Reappearance of PCR positivity with monitoring, if confirmed, should prompt a referral for allogeneic HSCT, if a potential candidate, as per above.

CNS prophylaxis
1. Intrathecal chemotherapy: All patients should receive intrathecal chemotherapy prophylaxis, starting with the initiation of induction therapy, and continuing through maintenance therapy, for 11-12 total doses.
   a. If the initial CSF is positive, intrathecal chemotherapy should be administered twice weekly until CSF clearance is confirmed on at least 3 occasions.
2. Cranial Radiation: Cranial radiation may be omitted from CNS prophylaxis, unless there is evidence of fixed CNS disease.

Supportive care
1. Antimicrobial prophylaxis against PJP and HSV/VZV should be used from induction through the completion of maintenance therapy. Antifungal prophylaxis should be used during intensive induction therapy.
2. Asparaginase therapy requires close monitoring for liver toxicity, pancreatitis, hypertriglyceridemia and VTE.
   a. Anticoagulation prophylaxis should be considered during intensification therapy. Documented VTE should be treated with full-dose anticoagulation but asparaginase therapy can generally be resumed after several weeks if stable or improved.
   b. Asparaginase therapeutic drug monitoring should be routinely performed in all patients at the end of the first intensification cycle, at a later cycle and after any dose reduction, targeting a trough level of \( \geq 0.1 \text{ U/L} \).
   c. Dose reductions should be considered for grade 2 or greater hepatic toxicity.
   d. Pancreatitis requires discontinuation of asparaginase.
3. Prophylaxis against corticosteroid-induced osteoporosis and gastritis should be used.

Relapsed ALL
1. Transplant-eligible BCR-ABL negative B-ALL patients should be treated with an antibody-based regimen, either blinatumomab or inotuzumab (if CD22 positive).
2. BCR-ABL positive patients relapsing on an imatinib-based regimen should be treated with a second generation or third generation TKI + either chemotherapy, inotuzumab or blinatumomab. If a T315I mutation is detected, ponatinib plus blinatumomab or chemotherapy should be used.
3. T-ALL patients should receive a non-cross resistant re-induction chemotherapy regimen, or nelarabine.
4. Transplant eligible patients should be considered for allogeneic HSCT in CR-2, if not performed in CR-1.

CAR T-Cell Therapy:
1. Indicated for fit B-ALL patients relapsing after allogeneic HSCT, refractory to 2 induction regimens, or relapsed and not considered suitable candidates for HSCT.
2. Patients should be referred for apheresis prior to administering salvage immunosuppressive chemotherapy such as corticosteroids or cyclophosphamide, to avoid interfering with the quality of the product.
3. Patients should in most cases receive bridging/cytoreductive therapy following apheresis, to prevent clinical deterioration due to disease progression and to achieve cytoreduction prior to CAR T infusion. This may consist of either chemotherapy or antibody-based therapy.
Discussion and Recommendations

Pathogenesis
ALL is thought to arise from interactions between exogenous or endogenous exposures, genetic susceptibility, and chance. Infection was the first suggested causal exposure for childhood acute lymphoblastic leukemia. After the Hiroshima and Nagasaki atomic detonations, ionizing radiation quickly became established as an exposure leading to childhood ALL. Chromosomal translocations occurring in utero during fetal hematopoiesis have been suggested as the primary cause for pediatric ALL while postnatal genetic events are considered secondary contributors. Many of these chromosomal rearrangements disrupt genes that regulate normal haematopoiesis and lymphoid development (e.g. RUNX1, ETV6), activate oncogenes (e.g. MYC), or constitutively activate tyrosine kinases (e.g. ABL1). Patients with trisomy 21, Klinefelter’s syndrome and inherited diseases with excessive chromosomal fragility such as Fanconi’s anemia, Bloom’s syndrome and ataxia-telangiectasia have a higher risk of developing ALL. However, in the majority of ALL patients no gross chromosomal alteration is noted suggesting that additional submicroscopic genetic alterations likely contribute to leukaemogenesis. Genome-wide association studies of childhood have noted common allelic variants in IKZF1, ARID5B, CEBPE, and CDKN2A which have been significantly and consistently associated with childhood ALL. Others have investigated the associations of genetic polymorphisms in folate pathway and DNA repair genes with susceptibility to ALL. Although several genetic alterations have well established roles in leukemogenesis (e.g. activating mutations in NOTCH1) the roles of many others remains elusive.

Clinical Manifestations, Diagnosis, and Work-up
Clinical manifestations of ALL are highly variable. At presentation patients may have a multitude of constitutional symptoms, easy bruising, bleeding, dyspnea, dizziness, and infections due to anemia, thrombocytopenia and neutropenia. Extremity and joint pain may be the only presenting symptoms in some patients. Lymphadenopathy, splenomegaly and/or hepatomegaly are seen on physical examination in approximately 20% of patients. Abdominal masses from gastrointestinal involvement or chin numbness from cranial nerve involvement may be seen but are more suggestive of mature B-ALL. Less than 10% of patients have symptomatic central nervous system (CNS) involvement. T-lineage ALL with a mediastinal mass can cause stridor and wheezing, pericardial effusions, and superior vena cava syndrome. Testicular involvement is rare in adults.

The diagnosis of ALL begins with an evaluation of the peripheral blood film which may identify the presence of blasts. Patients presenting with only a mediastinal mass or lymphadenopathy require a tissue biopsy. All patients should have a bone marrow examination. As per the 2008 and 2016 WHO classifications, in contrast to myeloid malignancies, there is no agreed-upon lower limit for the percentage of blasts required to establish a diagnosis of lymphoblastic leukemias. Despite this, some guidelines suggest that the diagnosis of ALL requires demonstration of > 20% blasts in the bone marrow aspirate/biopsy material. By convention the term lymphoma is used when the process is confined to a mass lesion with no or minimal evidence of peripheral blood and bone marrow
involvement. Based on morphologic, genetic and immunophenotypic features, lymphoblastic lymphoma is indistinguishable from ALL, and is in fact not distinguished in the WHO 2016 classification.

The assessment of immunophenotype by flow cytometry is essential to establishing the diagnosis. The initial immunophenotyping panel should be comprehensive enough to establish a leukemia associated phenotype (LAP) to allow for use in minimal disease monitoring (MRD). Cytophenetic examination with examination of metaphases and/or fluorescence in situ hybridization (FISH) is crucial (eg, for BCR-ABL and MLL-AF4) particularly in cases in which cytogenetics are unavailable or have failed. Screening by polymerase chain reaction (PCR) for the BCR-ABL transcripts is also essential as it significantly impacts treatment. Determination of the BCR-ABL breakpoint is also required for subsequent molecular monitoring. Some labs also evaluate blast cells with molecular methods for the detection of patient-specific immunoglobulin and T-cell receptor (Ig/TCR) rearrangements; however, this is not routinely performed within Alberta. If bone marrow transplant is a consideration, tissue typing of both the patient and siblings should also be performed at diagnosis. If there are no HLA-matched siblings, consideration should be given to prompt initiation of an unrelated donor search.

**Recommendations:**
The initial work-up of patients with ALL should include a thorough history and physical examination as well as baseline laboratory investigations including complete blood count, chemistry with extended electrolytes, tests of renal and liver function including amylase and lipase, a disseminated intravascular coagulation panel, and a tumour lysis panel. Cardiac imaging e.g. echocardiogram, multigated acquisition (MUGA) scan or cardiac MRI should be undertaken for all patients due to the use of anthracyclines.

Bone marrow studies or peripheral blood studies incorporating, as noted above, immunophenotyping, cytogenetics, and molecular studies should be completed. Ideally results of BCR-ABL testing should be available prior to the initiation of induction therapy, and should be available within 5 days. Lastly, patients eligible for allogeneic stem cell transplantation, and their siblings should have human leukocyte antigen (HLA) typing performed.

**Classification and Prognostication**

**Classification of ALL:**
The WHO 2016 classification is largely based on recurrent cytogenetic abnormalities, but also includes molecular markers such at KMT2A and BCR-ABL (Table 1). There is also a new provisional entity encompassing the BCR-ABL-like genotype (described later), which is not to this point readily diagnosed using routine testing. B-lymphoblastic leukemia/lymphoma NOS encompasses all subtypes not otherwise defined by one of the recognized abnormalities.
It is notable that this classification does not primarily distinguish between different immunophenotypic features, apart from the provisional T cell entities. It also does not distinguish between T-lymphoblastic lymphoma and T-ALL, acknowledging the widely held view that these are different clinical presentations of the same disease, and should be managed similarly. On the other hand, early T-cell precursor (ETP) lymphoblastic leukemia has been identified as a distinct subtype with unique biologic and clinical characteristics.

Table 1: WHO (2016) classification of ALL

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<th>B-lymphoblastic leukemia/lymphoma</th>
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<tr>
<td>B-lymphoblastic leukemia/lymphoma, NOS</td>
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<td>B-lymphoblastic leukemia/lymphoma with recurrent genetic abnormalities</td>
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<tr>
<td>B-lymphoblastic leukemia/lymphoma with t(9;22)(q34.1;q11.2);BCR-ABL1</td>
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<tr>
<td>B-lymphoblastic leukemia/lymphoma with t(v;11q23.3);KMT2A rearranged</td>
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<tr>
<td>B-lymphoblastic leukemia/lymphoma with t(12;21)(p13.2;q22.1);ETV6-RUNX1</td>
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<td>B-lymphoblastic leukemia/lymphoma with hyperdiploidy</td>
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<tr>
<td>B-lymphoblastic leukemia/lymphoma with hypodiploidy</td>
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<tr>
<td>B-lymphoblastic leukemia/lymphoma with t(5;14)(q31.1;q32.3) IL3-IGH</td>
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<tr>
<td>B-lymphoblastic leukemia/lymphoma with t(1;19)(q23;p13.3);TCF3-PBX1</td>
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<tr>
<td>Provisional entity: B-lymphoblastic leukemia/lymphoma, BCR-ABL1–like</td>
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<tr>
<td>Provisional entity: B-lymphoblastic leukemia/lymphoma with iAMP21</td>
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<th>T-lymphoblastic leukemia/lymphoma</th>
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<tr>
<td>Provisional entity: Early T-cell precursor (ETP) lymphoblastic leukemia</td>
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<tr>
<td>Provisional entity: Natural killer (NK) cell lymphoblastic leukemia/lymphoma</td>
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Prognostic Factors in ALL:
Prognostic models for ALL have been refined continuously with improvements in therapy rendering some prognostic variables invalid. The following represent clinical, cytogenetic and important molecular risk factors. It is important to note that many of these factors were defined using adult-based protocols and more studies are needed to evaluate their significance with pediatric based protocols.

Clinical Prognostic Factors:

Age, Gender and Ethnicity
Age is an important prognostic factor in ALL. In children age (infant or ≥10 years old) is an unfavourable risk group especially those younger than 6 months old. Regardless of the treatment protocol utilized older adults are regarded as a prognostically unfavorable group. One study noted that the outcomes of patients aged over 55 had a probability of survival of 20% at 3 years while others have noted that patients over the age of 35 have poorer outcomes. Recent data suggest that adolescents and young adults benefit with improved overall survival if treated according to
pediatric protocols. Several studies have suggested that the influence of age may relate to the increased prevalence of poor-risk features while others have suggested that it is independent of cytogenetic and molecular aberrations. The ability to tolerate chemotherapy likely plays an important role. Together with age, male gender and race (Hispanic or of African descent) have been considered negative prognostic factors in pediatric ALL Inaba, 2013 #20. Racial differences in prognosis have been linked to socioeconomic factors but also to differences in genomic variations. For example, germline single nucleotide polymorphisms of PDE4B and ARID5B are associated with Native American genetic ancestry and somatic CRLF2 were over-represented in children with a Hispanic background.

**White Blood Cell Count**

In children, a presenting leucocyte count (≥50 × 10⁹/L) has been associated with a worse prognosis. In many, but not all adult studies of ALL, high-risk ALL has been defined as WBC ≥ 30 x 10⁹/L for B-cell ALL and ≥100x10⁹/L for T-cell ALL. Although most early studies identifying this factor used adult-based protocols, subgroup analyses in larger studies show that these continue to be important prognostic factors with pediatric-inspired regimens Brandwein, 2011 #51;Boissel, 2003 #357. In their study of the DFCI protocol in adults a high WBC (>30 x 10⁹/L for Pre-B ALL or >100 x 10⁹/L for T-ALL) was associated with inferior RFS and OS. However, subsequent studies incorporating early MRD detection into multivariate models have demonstrated that baseline WBC was no longer an independent predictor of relapse (see section on MRD).

**Immunophenotype**

The SEER database demonstrated a better prognosis with B cell as compared with T cell immunophenotype in patients < age 20 years; while in patients ≥ 20 years of age, T cell immunophenotype was more favorable. This was confirmed in a metanalysis by Kako et al. of published studies from January 1998 to March 2013 to compare the outcomes of chemotherapy for T- and B- lineage ALL and noted superior survival in patients with T-lineage ALL compared to those with B-lineage ALL although the inclusion of patients with Ph+ ALL likely influenced the.

Amongst adults T-cell ALL accounts for 14–22% of adult ALL and is thought to be of favourable prognosis. In both the LALA 87 and the UKALL/EGOG 2993 trial, T-ALL was associated with male gender, age <35 - 39 years old, CNS involvement and a high WBC count. The LALA-87 investigators also noted a higher incidence of a mediastinal mass and anemia. In this study for patients age <40 treated with chemotherapy alone 3 year DFS was superior in the group with a T-cell phenotype relative to a B-ALL phenotype (59% vs. 20%). No difference in DFS was seen in patients with B- or T-ALL patients treated with allo- or auto – HSCT. Similarly, Both Rowe et al. and Larson et al. noted that improved OS in patients with T-cell antigen expression relative to B-lineage antigens. Kantarjian et al. using Hyper-CVAD noted similar results. Using the pediatric Dana Farber Cancer Institute (DFCI) protocol a trend toward improved clinical outcomes was observed in adolescents and adults with T-ALL. The GRAALL study group noted that when using a pediatric protocol at
42 months, EFS was estimated to be 62% (95% CI, 50% to 72%) in T-ALL patients and 52% (95% CI, 42% to 59%) in BCP-ALL patients (p=0.09). Within the T-cell ALL subset the prognosis is worse for pro-, pre- and mature-T subtypes (CD1a-, CD3-/CD3+) compared with the CD1a+ cortical/thymic phenotype. The early T-cell precursor ALL which retains stem cell-like features is associated with a dismal prognosis with conventional chemotherapy\(^\text{27}\) in both adult and pediatric T-ALL.\(^\text{28}\)

As noted above, several studies have suggested that patients with B-cell phenotype fare worse than those with T-ALL. Patients with a CD10-negative pro-B phenotype are considered as high-risk particularly when associated with t(4;11)/abn q23.\(^\text{1, 26}\) The pre-B subtype expressing cytoplasmic heavy chains has a bad outlook when harboring MLL rearrangements. The CD20 antigen is expressed in nearly half of B-cell ALL and its impact on clinical outcomes is controversial. Maury et al. when using the pediatric GRAALL 2003 protocol in adults aged 15-60 years old with Ph-negative ALL noted that CD 20 expression did not influence achievement of complete remission but was associated with a higher cumulative incidence of relapse (CIR) and lower EFS at 42 months (42% vs. 29%) in patients with a WBC ≥ 30 ×10⁹/L (P=0.006). Thomas et al. also noted an inferior survival in CD20 positive patients using the adult-based hyper-CVAD protocol.\(^\text{29}\) In contrast, a retrospective analysis by the Princess Margaret Hospital groups in adult patients, most of whom had received a pediatric-based regimen, did not find any association between CD20 expression and outcome\(^\text{30}\) and in-fact there appeared to be a trend towards a favourable EFS in those showing CD20 positivity.

**Cytogenetic Studies**

Karyotype is an important prognostic factor with a number of cytogenetic abnormalities being associated with altered prognosis in ALL (Table 2). The frequency of cytogenetic aberrations varies between adult and childhood ALL and may partially explain the differences in clinical outcomes between patient populations.\(^\text{8}\) Whether cytogenetic abnormalities remains important with the use of pediatric protocols in adult patients remains unclear; some studies suggest that most abnormalities are not independent predictors of outcome in adults treated with such protocols, with the exception of KMT2A-associated abnormalities.\(^\text{20, 21, 26}\)

The Philadelphia chromosome, characterized by the t(9;22) translocation resulting in production of a BCR-ABL1 fusion gene and protein, is the most common cytogenetic and molecular abnormality in adult ALL. The frequency is age-dependent, being present in approximately 8–10% of adolescents, 15-30% of younger and middle aged adults, and 40-50% of elderly ALL patients.\(^\text{9}\) Over two-thirds have the p190 gene, with the remaining harbouring the p190 gene.

Until recently the BCR–ABL1 fusion gene marked the most unfavourable subgroup of adult ALL. With chemotherapy alone the CR rate was 75%- 80%, median DFS about 10 months and 5-year survival below 10-20%.\(^\text{26, 31, 32}\) Most studies demonstrated superior outcomes with allogeneic HSCT compared to chemotherapy alone.\(^\text{26, 31, 32}\) Combinations of tyrosine kinase inhibitors (TKIs) with chemotherapy have produced superior outcomes to chemotherapy alone, as will be discussed later.
The t(4;11) is present in up to 60% of infants younger than 12 months but is uncommon in adult patients, constituting 5-10% of cases. When rearranged, the MLL (now called KMT2A) gene has been found to be associated with inferior RFS and OS in adult patients when using a pediatric protocol. The t(1;19) is uncommon in adults; its prognostic significance is unclear, with conflicting data. Garg et al. noted that adults treated with Hyper CVAD had a significantly better CRD and OS compared with all other patients. However, Foa et al. found that this abnormality is frequently associated with early treatment failure, and recommended that these patients should be considered for intensified treatment strategies. The t(12;21) abnormality leading to ETV6-RUNX1 fusion is detectable in about 25% of children and 3% of adults with B-ALL. Patients generally have a favorable prognosis.

Hyperdiploidy (>50 chromosomes) is seen in approximately 25% - 30% of pediatric cases and 7% of adults and represents the most common chromosomal abnormality in children. It is associated with a favourable prognosis regardless of age and leukocyte count at presentation. Its characteristic genetic feature is the nonrandom gain of chromosomes X, 4, 6, 10, 14, 17, 18 and 21, with individual trisomies or tetrasomies being seen in over 75% of cases. The individual structural abnormalities do not appear to influence outcome in patients with hyperdiploidy except for the t(9;22), which is associated with a poor prognosis. The favourable prognosis may reflect an increased propensity of these cells to undergo apoptosis. In contrast, 5% to 6% of ALL patients, independent of age, have the loss of various chromosomes, resulting in a hypodiploid clone with fewer than 44 – 46 chromosomes. These patients generally have a poor prognosis, especially those with near-haploid and low-hypodiploid clones. Recent data suggest that complex karyotypes (≥ five chromosomal abnormalities) occur more frequently with increasing age and may be associated with inferior survival.

Intrachromosomal amplification of chromosome 21 (iAMP21) occurs at an incidence up to 2% in older children with B-cell precursor ALL. Intrachromosomal amplification of chromosome 21 (iAMP21) occurs at an incidence up to 2% in older children with B-cell precursor ALL. Harrison, 2005 #1056 and is defined by at least three copies of the RUNX1 gene (Children's Oncology Group, COG, definition for iAMP21). iAMP21 has been shown to be linked to a dismal outcome when patients are treated with standard therapy, because it is associated with an increased risk of both early and late relapses. It is rare in adults.

Many of these prognostic factors may be regimen dependent. Many older studies used adult-based protocol. In contrast, using pediatric based regimens, most of these cytogenetic abnormalities have not been found to be of prognostic importance in adults, with the notable exception of t(4;11).

Molecular Studies
Molecular genetics has identified several gene mutations, translocations and amplifications that may have prognostic significance in ALL. IKZF1 encodes IKAROS which has been established as one of
the most clinically relevant tumor suppressors in ALL. It is a DNA-binding zinc finger transcription factor that regulates the development and function of the immune system and acts as a master regulator of normal hematopoietic differentiation and proliferation, particularly in lymphoid lineage.\textsuperscript{48} Deletion of a single IKZF1 allele or mutations of a single copy of IKZF1 were first detected in 15\% of pediatric B-cell ALL and in more than 80\% of Ph+ ALL cases, either de novo Ph+ ALL or chronic myeloid leukemia at progression to lymphoid blast crisis suggesting a critical role in the pathogenesis of Ph+ ALL.\textsuperscript{49-52} The incidence in adults is about 50\% in B-cell ALL and about 65\% when BCR-ABL positive. Recent data further suggest that together with Ph+ve ALL, mutations in IKZF1 are also a hallmark of 'BCR–ABL1-like ALL'.\textsuperscript{5} Alteration of the IKAROS gene is associated with increased risk of treatment failure and relapse in both BCR–ABL1-positive and BCR–ABL1-negative disease independent of commonly used risk stratification features such as age, sex, white cell count, and levels of minimal residual disease (MRD).\textsuperscript{21, 52}

PAX5 encodes a transcription factor known as B-cell specific activator protein, that plays a key role in B-cell commitment by activating essential components of the B-cell receptor signaling and repressing the transcription of genes necessary for T-lymphopoiesis.\textsuperscript{53} Monoallelic deletion of PAX5 have been observed in about 30\% of children and adults\textsuperscript{54, 55} and have been demonstrated to not influence treatment outcome.\textsuperscript{36, 55, 56} The CDKN2A/B locus encodes for the INK4-class cyclin dependent kinase inhibitors p15INK4B, p16INK4A and for p14ARF. CDKN2A and CDKN2B deletions have been identified in 29\% and 25\% of BCR-ABL1-positive ALL patients, respectively.\textsuperscript{57} The association with prognosis is still controversial.\textsuperscript{36, 57, 58}

Four independent groups in late 2009 and early 2010 identified that up to 50\% of BCR-ABL1-like ALL cases have dysregulated expression of CRLF2, the gene encoding the cytokine receptor-like 2 factor.\textsuperscript{56, 59-61} Overall aberrant expression of CRLF2 was found in 12.5\% to 15\% of B-ALL that lacks typical chromosomal rearrangements and in 50–60\% of Down Syndrome (DS) associated ALL, suggesting that CRLF2 overexpression is especially relevant to tumorigenesis in patients with trisomy 21\textsuperscript{Mullighan, 2009 #1021;Hertzberg, 2010 #1030;Yoda, 2010 #1031}. Patients with CRLF2 rearrangements had extremely poor treatment outcomes compared with those without CRLF2 rearrangements (35.3\% vs 71.3\% relapse-free survival at 4 years).\textsuperscript{62}

Mutations of NOTCH1, a transmembrane receptor-encoding gene that regulates normal T-cell development, have been detected in in about 60\% of T-ALL.\textsuperscript{63} Early studies in paediatric T-ALL showed that NOTCH1 mutations may be associated with a favourable prognosis.\textsuperscript{21, 63-68} Similarly, in adult T-ALL patients, studies have demonstrated a better prognosis for patients with NOTCH1 and/or FBXW7 mutations,\textsuperscript{67, 69} but this could not be validated in a series of 88 patients treated in the MRC UKALLXII/ECOGE2993 protocol\textsuperscript{70} or by the Zhu et al who in fact noted that Chinese adult TALL patients with mutated NOTCH1 had poorer survival compared with those with wild-type NOTCH1.\textsuperscript{71} Overall these studies seem to suggest that NOTCH activation is associated with improved early
therapeutic response. However, this early benefit translates into improved overall survival only in some series, probably due to differences in therapy.\textsuperscript{63}

**Gene Expression Profiling:**

**BCR-ABL like ALL**
Mullighan and colleagues identified some patients with Ph-negative ALL which had a gene expression profile almost identical to Ph+ ALL and which were termed Ph-like ALL.\textsuperscript{54, 56, 72} This gene-signature has been noted in approximately 15% of pediatric cases, but the frequency is as high as 33% in adults with B-ALL.\textsuperscript{73} Genetic alterations of activating kinases or cytokine receptor signaling, including ABL, JAK2, CSF1 and EPOR, are commonly observed in addition to overexpression of cytokine receptor-like factor 2 (CRLF2) and frequent deletions of \textit{IKZF1}.\textsuperscript{54, 56, 72, 74} Some studies have found higher levels of MRD after induction therapy,\textsuperscript{74, 75} increased relapse rates and inferior overall survival.\textsuperscript{8} However, reliable testing for this genotype is not routinely available in most laboratories, including in Alberta.

**Early T-cell precursor acute lymphoblastic leukaemia (ETP – ALL)**
ETP – ALL accounts for 12% of paediatric T-ALL. It is an aggressive leukaemia characterised by an immature immunophenotype with lack of CD1a and CD8 expression, weak CD5 expression\textsuperscript{76} and aberrant expression of myeloid and stem cell markers (CD117, CD34, HLA-DR, CD13, CD33, CD11b, or CD65) on at least 25% of lymphoblasts.\textsuperscript{8} The long-term response to therapy is one of the worst among recognized high-risk forms of childhood ALL.\textsuperscript{27} In one study, the 10 year OS was 19% compared with 84% in the non-ETP ALL.\textsuperscript{27} Similarly inferior outcomes have been reported in adults with ETP-ALL.\textsuperscript{77, 78} The mutational spectrum in ETP-ALL is similar to myeloid tumours with a high frequency of activating mutations in the cytokine receptor and RAS signaling pathways including NRAS, KRAS, FLT3, IL7R, JAK3, JAK1, SH2B3, and BRAF, raising the possibility that addition of myeloid-directed therapies might improve the outcome of ETP ALL.\textsuperscript{79} Given the poor outcomes with conventional ALL regimens, most ETP-ALL patients are referred for allogeneic HSCT in CR1, where there are data suggesting a survival benefit.\textsuperscript{78}

**Other Factors:**

**Pharmacogenetics and pharmacogenomics**
Genome-wide analyses have identified specific gene signatures which may be prognostically relevant when associated with drug resistance e.g. polymorphism of genes metabolizing thiopurines, methotrexate, and cytarabine all of which have been associated with variable treatment response and are mechanisms of drug resistance.\textsuperscript{1} Rocha et al noted that the glutathioneS-transferasel1 (GSTM1) non-null genotype was associated with a higher risk of recurrence, which was increased further by the thymidylate synthetase (TYMS) 3/3 genotype. Others have observed that hyperdiploid cells
accumulate more methotrexate polyglutamates as they possess extra copies of the gene encoding reduced folate carrier, an active transporter of methotrexate. Hareedy et al found significant associations between variants in genes coding for enzymes and transporters related to the 6-mercaptopurine pathway and clinical outcomes as well as hematological toxicity (neutropenia, agranulocytosis and leukopenia) in pediatric patients with acute lymphoblastic leukemia.\textsuperscript{80} The membrane transporter P-glycoprotein, encoded by the ABCB1 gene, influences the pharmacokinetics of anti-cancer drugs. Gregers et al. noted statistically significant association between ABCB1 polymorphisms, efficacy and toxicity in the treatment of ALL.\textsuperscript{81}

**Table 2: Prognostic Factors in ALL**

<table>
<thead>
<tr>
<th>Abnormality</th>
<th>Clinical Impact</th>
<th>Notes</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Cyto-Genetics</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Philadelphia Chromosome</td>
<td>Poor prognostic indicator</td>
<td>8-10% of adolescents. 15-30% adults. 50% elderly.</td>
</tr>
<tr>
<td>MLL Rearrangements</td>
<td>Poor prognostic indicator</td>
<td>Significance in adult patients using paediatric protocols? Immature immunophenotype, B-cell lineage, co-expression of myeloid antigens and high leukocyte counts. 30% of childhood ALL; Adverse prognosis can be overcome with intensive chemotherapy in adults and children, increased risk of CNS relapse Jeha, 2009 #190. Adults treated with Hyper CVAD had better CRD and OS compared with all other patients Garg, 2009 #861.</td>
</tr>
<tr>
<td>t(1;19) [TCF3-PBX1]</td>
<td>Variable</td>
<td>30% of childhood ALL; Adverse prognosis can be overcome with intensive chemotherapy in adults and children, increased risk of CNS relapse Jeha, 2009 #190.</td>
</tr>
<tr>
<td>t(12;21)[ETV6- RUNX 1]</td>
<td>Favourable prognosis</td>
<td>Detectable in about 18-25% of children and 1-3% of adults.</td>
</tr>
<tr>
<td>iAMP of chromosome 21</td>
<td>Poor prognosis</td>
<td>2% of older children with B-ALL.</td>
</tr>
<tr>
<td>Hyperdiploidy</td>
<td>Favourable prognosis</td>
<td>25-30% of cases; nonrandom gain of chromosomes X, 4, 6, 10, 14, 17, 18 and 21; The favourable prognosis may reflect an increased propensity of these cells to undergo apoptosis.</td>
</tr>
<tr>
<td>Hypodiploidy</td>
<td>Poor prognosis</td>
<td>5-6% of ALL patients; near haploid and low-hypodiploid have the worst prognosis.</td>
</tr>
<tr>
<td>Complex Karyotype</td>
<td>Poor prognosis</td>
<td>More than 5 chromosomal abnormalities.</td>
</tr>
<tr>
<td><strong>Molecular Genetics</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>IKZF1 mutations</td>
<td>Poor prognosis</td>
<td>Most commonly present in BCR-ABL + ALL or BCR-ABL like ALL.</td>
</tr>
<tr>
<td>Abnormality</td>
<td>Clinical Impact</td>
<td>Notes</td>
</tr>
<tr>
<td>------------</td>
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<td>-------</td>
</tr>
<tr>
<td>PAX5</td>
<td>No effect on treatment outcomes</td>
<td>30% of adults and paediatric patients.</td>
</tr>
<tr>
<td>CDKN2A/B</td>
<td>Prognosis uncertain</td>
<td>CDKN2A – 29% of BCR-ABL positive patients CDKN2B – 25% of BCR-ABL positive patients</td>
</tr>
<tr>
<td>CRLF2</td>
<td>Extremely poor outcomes.</td>
<td>12.5% - 15% of B-ALL in patients lacking typical rearrangements; 5060% down syndrome ALL. 50% concomitant mutations in JAK ( \frac{1}{2} ).</td>
</tr>
<tr>
<td>NOTCH</td>
<td>Improved prognosis</td>
<td>Most common alteration in TALL; 60% of T-ALL Maybe associated with FBXW7 mutations – which may worsen survival.</td>
</tr>
</tbody>
</table>

**Molecular Profiling**

| BCR-ABL Like ALL | Poor prognosis | 15% of B-cell ALL. May be responsive to treatment intensity |
| Early T-cell phenotype | Poor Prognosis | Distinct gene-expression profile. Lack of CD1a, CD8 and weak CD5 expression. Long-term response to therapy one of the worst in childhood ALL. |

**Other Prognostic Factors**

| Epigenetics | Unknown prognostic significance | Up to 80% of patients; Unclear clinical implications. T-ALL expressing EZH2 had a lower probability of DFS compared to T-ALL negative for EZH2 Determine response to drugs and toxicities of ALL therapy. GlutathioneS-transferase 1 phenotype associated with higher risk of recurrence. Hyperdiploid cells accumulate more MTX and have increased toxicity. Variants in genes encoding enzymes and transporters related to the 6-MP pathways influence toxicity. Variants of ABCB1 (P-glycoprotein) maybe associated toxicity and efficacy. |
| Pharmacogenomics | | |

**Pre-Treatment Risk Stratification**

Over the past twenty years there has been continued debate regarding the risk stratification of patients with ALL with different groups using the above clinical, immunophenotype, cytogenetics and molecular tests to variably group patients into those that are standard risk, high risk or very high risk of having a leukemia relapse. As discussed, many of these discrepancies may be related to the type of treatment.
reigmens used (adult vs pediatric-based), and whether MRD analysis is taken into consideration in risk stratification.

Recent studies have found that, using pediatric-based protocols, most cytogenetic abnormalities are not independent predictors except for certain abnormalities, such as KMT2A-based abnormalities, t(9;22) and possibly hypodiploidy.\textsuperscript{21, 45} Given the poor prognosis of patients with Ph+ve ALL, the initial risk stratification for all patients should be based on the presence or absence of t(9;22)/BCR-ABL1. Amongst Ph-ve patients the NCCN considers those with hypodiploidy (<44 chromosomes), t(v;11q23)/KMT2A rearrangements or complex karyotype (≥5 chromosomal abnormalities) as high risk.\textsuperscript{8}

Although prior studies have shown that a WBC > 30 for B-cell ALL and >100 for T-cell ALL were important risk factors,\textsuperscript{20} MRD has since been demonstrated to be a more important predictor on multivariate analysis (discussed below). However, age remains an important predictor of outcome in nearly all studies, with patients > age 30-35 having worse outcomes than so-called AYA (adolescent and young adult) patients.\textsuperscript{20}

Recent evidence suggests that molecular profiling, and specifically the detection of a Ph-like genetic signature as discussed, may be the most important pre-treatment predictor of outcome in adults and children with BCR-ABL negative B-ALL. However, detection is time consuming and labour intensive, and is not routinely available at most centres, including in Alberta.

**Table 3: Pre-treatment risk stratification factors in ALL patients treated with 'adult' chemotherapy regimens.\textsuperscript{1}**

<table>
<thead>
<tr>
<th>Study Group</th>
<th>Age</th>
<th>WBC</th>
<th>Immunophenotype</th>
<th>Cytogenetics</th>
<th>Others</th>
<th>Definition of Risk Group</th>
</tr>
</thead>
<tbody>
<tr>
<td>CALGB\textsuperscript{82}</td>
<td>&gt;60</td>
<td>&gt;30</td>
<td>Ph+</td>
<td>t(4;11)</td>
<td>L3, Med</td>
<td>SR = 0-1 HR = 2-4</td>
</tr>
<tr>
<td>GIMEMA\textsuperscript{83}</td>
<td>&gt;30</td>
<td>&gt;50</td>
<td>Ph+</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>JALSG\textsuperscript{84}</td>
<td>&gt;30</td>
<td>&gt;30</td>
<td>Ph+</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>MDACC\textsuperscript{12}</td>
<td>-</td>
<td>&gt;50</td>
<td>Ph+</td>
<td></td>
<td>ECOG 3-4, L2, d14 BM+</td>
<td>SR = 0-1 IR = 1 HR = 2/Ph+</td>
</tr>
<tr>
<td>GOELAMS\textsuperscript{85}</td>
<td>&gt;35</td>
<td>&gt;30</td>
<td>B-Cell</td>
<td>Ph+, t(4;11), (t1;19)</td>
<td>CR &gt; 1c</td>
<td>SR = 0 HR1 = 1-2 HR2 = 2-3</td>
</tr>
<tr>
<td>LALA\textsuperscript{86}</td>
<td>-</td>
<td>&gt;30</td>
<td>Myeloid Markers</td>
<td>Ph+, t(4;11), 11q23, (t1;19)</td>
<td>CR &gt; 1c, CNS +</td>
<td>SR = 0</td>
</tr>
<tr>
<td>Study Group</td>
<td>Age</td>
<td>WBC</td>
<td>Immunophenotype</td>
<td>Cytogenetics</td>
<td>Others</td>
<td>Definition of Risk Group</td>
</tr>
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<td>-------------------------</td>
</tr>
<tr>
<td>PETHEMA</td>
<td>30-50</td>
<td>&gt;25</td>
<td></td>
<td>Ph+, t(4;11), 11q23, (t1;19)</td>
<td>SR = 0</td>
<td>HR ≥ 1, Ph+, CNS+</td>
</tr>
<tr>
<td>MR-ECOG</td>
<td>&gt;35</td>
<td>&gt;30 (B-Cell) &gt;100 (T-Cell)</td>
<td>Ph+</td>
<td></td>
<td>SR = 0</td>
<td>HR ≥ 1</td>
</tr>
<tr>
<td>HOVON</td>
<td>-</td>
<td>&gt;30 (B-Cell) &gt;100 (T-Cell)</td>
<td>Ph+, t(4;11), t(1;19) CR &gt; 4 weeks</td>
<td>SR = 0</td>
<td>HR ≥ 1</td>
<td></td>
</tr>
<tr>
<td>GMALL</td>
<td>15-55</td>
<td>&gt;30 (B-Cell) Pro-B Early/Mature T</td>
<td>Ph+, t(4;11) CR &gt; 3 weeks</td>
<td>SR = 0</td>
<td>MRD-HR ≥1, MRD + VHR = Ph+</td>
<td></td>
</tr>
<tr>
<td>NILG</td>
<td>-</td>
<td>&gt;30 (B-Cell) &gt;100 (T-Cell) Pro-B Early/Mature T</td>
<td>Ph+, t(4;11), adverse cytogenetics CR &gt; 1c</td>
<td>SR = 0</td>
<td>MRD – HR = MRD + VHR = Ph+, t(4;11)+</td>
<td></td>
</tr>
</tbody>
</table>

SR – Standard Risk; HR – High Risk; IR – Intermediate Risk; VHR – Very High Risk; CR – Complete Remission; ECOG – Eastern Cooperative Oncology Group; Ph – Philadelphia; 1c – 1 cycle of chemotherapy; BM – Bone Marrow; MRD – Minimal Residual Disease

**Table 4:** Pre-treatment risk stratification factors in ALL patients treated with ‘paediatric’ chemotherapy regimens.
<table>
<thead>
<tr>
<th>Study Group</th>
<th>Age</th>
<th>WBC</th>
<th>Immunophenotype</th>
<th>Cytogenetics</th>
<th>Others</th>
<th>Definition of Risk Group</th>
</tr>
</thead>
<tbody>
<tr>
<td>DFCI - 91 – 01⁹²</td>
<td>&lt;2 or &gt;9</td>
<td>&gt;20</td>
<td>T-cell</td>
<td>Ph+, Mediastinal mass, CNS leukemia,</td>
<td>SR = 0</td>
<td>HR ≥ 1</td>
</tr>
<tr>
<td>DFCI – 95 – 01⁹²</td>
<td>&lt;1 or &gt;10</td>
<td>&gt;50</td>
<td>T-cell</td>
<td>Ph+, Mediastinal mass, CNS leukemia,</td>
<td>SR = 0</td>
<td>HR ≥ 1</td>
</tr>
<tr>
<td>PETHEMA⁹³</td>
<td>&gt;30</td>
<td>-</td>
<td>t(9;22), t(1;19), t(4;11), MLL rearrangements.</td>
<td>SR = 0</td>
<td>HR ≥ 1</td>
<td></td>
</tr>
<tr>
<td>GRAALL⁹⁴</td>
<td>&gt;30 – B-cell</td>
<td></td>
<td>CNS involvement t(4;11) or MLL-AF4 fusion transcript, t(1;19) and/or E2A-PBX1 fusion transcript, low hypodiploidy (30 to 39 chromosomes or DNA index 0.85), near-triploidy (60 to 78 chromosomes or DNA index of 1.30 to 1.69). CsR and/or ChR, absence of CR after the first induction course, Ph+</td>
<td>SR = 0</td>
<td>HR ≥ 1</td>
<td></td>
</tr>
<tr>
<td>DFCI²⁰</td>
<td>≥35</td>
<td>&gt;30 B-Cell &gt;100 T-Cell</td>
<td>MLL</td>
<td>Not specified</td>
<td></td>
<td></td>
</tr>
<tr>
<td>DFCI⁴⁵</td>
<td>≥34</td>
<td>&gt;30 B-Cell &gt;100 T-Cell</td>
<td>Not specified</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>DFCI⁹⁵</td>
<td>≥35</td>
<td>≥35 B-Cell</td>
<td>MLL, Ph+,</td>
<td>Not specified</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

SR – Standard Risk; HR – High Risk; IR – Intermediate Risk; VHR – Very High Risk; CR – Complete Remission; ECOG – Eastern Cooperative Oncology Group; Ph – Philadelphia; 1c – 1 cycle of chemotherapy; BM – Bone Marrow

**Recommendations:**
All patients should be classified as having B-cell or T-cell ALL based upon immunophenotyping results. Although some of the above noted prognostic factors are beyond the scope of routine clinical laboratories, all patients, should undergo cytogenetic evaluation and, if unsuccessful, FISH for the determination of the most clinically significant abnormalities, in particular BCR-ABL and MLL gene rearrangements. There is no convincing evidence that other cytogenetic abnormalities or cell surface
markers add to risk-stratification when using paediatric or paediatric inspired protocols. Given the increasing recognition of minimal residual assessments, all patients should have immunophenotyping performed. This can be used to guide post-induction treatment.

**Post-Treatment Risk Stratification**

**Role of MRD Assessments in the Management of Patients with ALL:**

Measurable (or minimal) residual disease (MRD) refers to the detection of small amounts of residual disease, undetectable by morphology. Techniques for MRD detection include multiparameter flow cytometry (MPFC) or molecular techniques. For BCR-ABL+ ALL, molecular detection is readily performed by qRT-PCR for BCR-ABL1, and is regarded as the gold standard for MRD detection. For BCR-ABL negative ALL, many European studies have used immunoglobulin gene rearrangements for MRD detection; however, this technique is labour intensive and requires detection of the specific rearrangement for each patient. Consequently, MPFC is widely used for MRD detection due to its ease, and has a sensitivity of $10^{-3}$ – $10^{-4}$; it is the technique used in Alberta.

Based on the paediatric literature, a number of studies over the past 15 years have explored the role of MRD in adults with ALL. Most of these have suggested that MRD may be the single most important factor predicting clinical outcomes. A number of investigators have described differences in patterns and dynamics of clearance of MRD between adult and childhood ALL as well as between B- and T- cell ALL. Foroni et al. noted that MRD decreased faster in children than in adults particularly in the first 6 months of CR while Parekh et al. noted that MRD clearance was slower with T-ALL. Bruggemann and colleagues measured MRD at 9 different time points ranging from 11 days post-induction up to 52 weeks post induction. Only a minority of patients had undetectable MRD at day +11 while at 6 weeks approximately 50% had undetectable MRD.

Several groups have explored the prognostic value of MRD in adult ALL patients. Bruggman and colleagues noted that a combination of MRD measurements at day 11, day 24 and 16 weeks could classify patients into those with a low likelihood of ALL relapse (MRD negative at day 11 and day 24), high likelihood of relapse (MRD positive at week 16) or intermediate risk of relapse (all others). Similar findings were noted by Bassan and colleagues who measured MRD at week 10 and week 22 post induction chemotherapy. Regardless of whether they were SR, HR or VHR by traditional criteria, patients who were MRD negative had significantly better DFS relative to those that were MRD positive.

Whether treatment intensification can negate the negative effects of residual disease has also been investigated by several investigators. Bassan and colleagues assessed MRD at week, week 16 and week 22. Patients that were MRD negative or with unknown MRD status but standard risk by traditional criteria received maintenance treatment only while patients that were MRD positive or with unknown MRD but high risk or very high risk by traditional criteria received an allogeneic SCT or autologous blood stem cell harvest/reinfusion with subsequent maintenance therapy. MRD positive patients receiving either SCT or
intensive chemotherapy had improvements in DFS with no significant difference between those that received SCT and intensive chemotherapy. Moreover, patients who became MRD negative had improved DFS relative to those who remained MRD positive. Similar results were noted by Gokbuget et al. where 5 year CCR improved for MRD positive patients undergoing an allo SCT. Ribera and colleagues noted that allo SCT in MRD –ve patients was unnecessary and counterproductive as it led to worse DFS and overall survival both in the whole series and an intention to-treat-analysis.

Beldjord et al. assessed 423 adult patients with Ph-negative ALL treated with a pediatric protocol. MRD1 was evaluated at 6 weeks after induction initiation and MRD2 after the first consolidation phase i.e. 12 weeks after induction initiation. As expected, MRD2 and MRD1 levels strongly correlated in this cohort. Overall, 265 patients achieved a MRD response at MRD1, 57 achieved it at MRD2 only, and 49 did not achieve it at either time-point. At 5 years, CIR was estimated at 24.7% in patients who reached a MRD level lower than 10^{-4} at MRD1, while it was 56.0% in those who reached this level at MRD2 only or never reached it, respectively. The value of traditional risk factors was examined in this population of patients treated with a pediatric-inspired protocol in multivariable analysis, against MRD1 response. Based on multivariate analysis, MLL gene rearrangement, IKZF1 gene deletion, and MRD1 level ≥10^{-4} were significant factors in B-ALL patients whereas a high-risk genetic profile and MRD1 level ≥10^{-4} were significant in T-ALL patients. Based on these results, high-risk patients could be defined as patients with MRD1 level ≥10^{-4}; and/or unfavorable genetics, defined as t(4;11) translocation or other MLL gene rearrangement and/or IKZF1 gene deletion in B-ALL; and 2) no NOTCH1/FBXW7 mutation and/or N/K-RAS mutation and/or PTEN alteration in T-ALL patients. MRD level seemed to be the predominant predictor in B-ALL patients with the risk further refined by genetic features. In contrast, in T-ALL patients both the oncogenic classification and MRD response were the major predictors of relapse. In both lineages, patients with high-risk genetic characteristics and poor MRD1 response experienced a worse outcome.

A study by the GRAALL group, using a pediatric inspired protocol for adults with B-ALL, evaluated the role of allogeneic HSCT according to MRD response with induction therapy. Patient who were MRD positive, defined as a level > 10^{-3} following induction therapy, had a significantly higher relapse rate and inferior OS as compared to those who achieved a level < 10^{-3}. Furthermore, those with MRD levels > 10^{-3} who underwent subsequent HSCT in CR1 had a significantly superior RFS and OS as compared to those who were not transplanted. In contrast, those with MRD levels < 10^{-3} following induction did not benefit from HSCT. These effects were seen for both B-ALL and T-ALL.

The Edmonton group subsequently analyzed outcomes following DFCI induction therapy. Between 2013-2019 patients with BCR-ABL negative ALL underwent induction therapy with this protocol, and MRD post-induction was assessed by multiparameter flow cytometry, with a sensitivity of 0.1%. Of 46 patients who achieve CR, 26 (57%) were MRD negative and 43% were MRD positive. The cumulative incidence of relapse was 45% in patients who were MRD positive at a level of >0.1%, as compared with 12% in MRD negative patients (p=0.05) (unpublished data). These results essentially mirror those reported by the GRAALL group as described above.
These data further support the conclusion that patients who fail to achieve a 3 log reduction in MRD levels with intensive induction therapy represent a high-risk group for relapse, and that these patients should be considered for HSCT in CR1. In contrast, those who achieve who achieve a >3 log reduction with induction can be successfully managed by chemotherapy alone with a low relapse risk and favourable prognosis. A subsequent German study (Herold et al, 2017) found a strong association between MRD positivity and a Ph-like genotype; Ph like B-ALL patients only achieved a 33% MRD negativity rate, as compared with 79% for Ph negative and non-Ph-like B-ALL patients (p=0.02). Therefore, MRD may represent a surrogate marker for a more resistant disease biology which is more destined to relapse with conventional chemotherapy.

**Treatment of MRD positive B-ALL:**
Blinatumomab is a bispecific tumour-engaging (BiTE) antibody with an anti-CD19 domain that binds to B-cells, including B-ALL cells, and an anti-CD3 domain that engages T lymphocytes to lyse the B cells. A large European study was recently published, evaluating the role of blinatumomab in 116 B-ALL patients in hematologic CR with MRD positivity, defined as a level > 0.1% by qRT-PCR. Patients were permitted to received up to 4 treatment cycles, and could undergo allogeneic HSCT at any time after the first cycle. Overall, 78% of patients achieved an MRD negative state, with a sensitivity of $10^{-4}$, after one blinatumomab treatment cycle. Two additional patients achieved this after 2 cycles. Of patient in first CR, 83% achieved MRD negativity. The treatment was well-tolerated. The median OS of the patients who achieved MRD negativity was 38.9 months, vs. 12.5 months in those who did not achieve MRD negativity (p=0.002); corresponding RFS were 23.6 vs. 5.7 months, respectively (p=0.002). With a median follow-up of 24 months, 49% of patient who underwent subsequent HSCT remained in continuous CR, as compared with 25% who did not undergo HSCT. By comparison, in Edmonton BCR-ABL negative B-ALL patients who were MRD positive at a level > 0.1% by flow cytometry after DFCI induction therapy were treated with intensified chemotherapy, using cycles 1A and 1B of Hyper-CVAD, between 2013-2018. Of 13 patients, only 5 (38%) were able to achieve MRD negativity; treatment was associated with considerable toxicity, including severe myelosuppression and mucositis.

These data indicate that (1) blinatumomab is able to induce a high rate of MRD negativity, usually after one cycle, with good tolerance, (2) patients who achieve MRD negativity have superior outcomes compared with those who do not, (3) patients who subsequently undergo allogeneic HSCT have favourable outcomes as compared with those who do not, and (4) intensified chemotherapy is less effective at inducing MRD negativity but is associated with considerably more toxicity. The use of blinatumomab for MRD positive B-ALL has now been widely incorporated into standard ALL protocols in Europe and North America.

**Recommendations:**
Taken together, these data indicate that MRD assessment provides critical prognostic information in adults with ALL. There is convincing evidence in pediatric and adult ALL that a high level of MRD at the end of induction therapy is associated with a higher relapse rate. Furthermore, the persistence of
MRD during consolidation and maintenance therapy, or its re-emergence, all seem to herald relapse. In contrast, negative MRD results are associated with a favorable prognosis. It is therefore recommended that all patients have MRD ascertained at the end of intensive induction therapy, or early during intensification. Patients with persistent MRD at a level > 0.1% should receive further treatment with an intent to achieve MRD negativity; for B-ALL the treatment of choice is blinatumomab, while for T-ALL intensified chemotherapy would be appropriate. Furthermore, these patients should be referred for allogeneic HSCT if suitable candidates, optimally after achieving MRD negativity. In contrast, patient achieving MRD negativity do not appear to benefit from transplant, provided they can successfully complete the subsequent treatment protocol.

**Treatment Approaches in BCR-ABL negative ALL**

In general, the treatment of ALL is complex consisting of several different chemotherapy cycles and, for some patients, stem-cell transplantation. A number of different approaches have been used as discussed below.

**Adult Multidrug Regimens:**

Starting in the 1960s, researchers at St. Jude Children’s Research Hospital designed combination therapies of available anti-leukemia drugs that were delivered in a sequence of extended courses of therapy. Since then several multidrug combinations have been developed centered on a vincristine, prednisone, and anthracycline combination, with or without asparaginase and cyclophosphamide. This concept was modified in children to the Berlin-Frankfurt Munster (BFM) ALL model and later, by Hoelzer et al., to adult ALL. Studies using this approach are presented in Table 5. Despite variations in chemotherapy regimens, variable use of allogeneic SCT or auto SCT the 5 year overall survivals ranged from 35 – 60% in various subgroups with induction mortality ranging of 5-10%.

The second treatment model pioneered at the M.D. Anderson Cancer Center consists of hyperfractionated cyclophosphamide, vincristine, doxorubicin, and dexamethasone, alternating with high-dose methotrexate and cytarabine (Hyper–CVAD). The regimen consists of a total of eight courses: four courses of hyper-CVAD (courses 1, 3, 5, and 7) alternate with four courses of MTX and HIDAC (courses 2, 4, 6, and 8). In the original report the mortality was 6% and the overall CR rate was 91%. The estimated median survival time was 35 months with a 5-year estimated survival of 39%. Younger age was associated with better survival (age <30 had estimated 5-year survival rate of 54% vs. age > 60 estimated 5-year survival rate of 25%). In an updated report of patients aged 15 – 92 years old, the CR rate for patients age ≤30 years was 99% and for patients age ≥60 years 80%, mostly because of a higher induction mortality. Kantarjian, 2004 #710. The addition of rituximab was associated with CCR and OS rates of 60% and 58%, respectively. Treatment with Rituximab–hyper-CVAD was associated with improvement in 3-year CRD rates compared with hyper-CVAD (78% vs. 53%). In patients age < 60 improvements in 3-year OS (75% vs47%; P 0=.003) rates favoring rituximab were observed.

1
The Pediatric Approach:
A third approach was the adoption of pediatric protocols or “pediatric inspired” regimens particularly for adolescents and young adults variably defined as patients 15 to 35-45 years old.1,106 These regimens have common features, including significantly increasing the non-myelosuppressive agents such as vincristine and steroids, using much higher cumulative doses of asparaginase for prolonged asparagine depletion, and administering very early and extended intrathecal methotrexate together with high-dose systemic methotrexate. Several studies from Europe and the USA reported that pediatric inspired approaches are feasible in adolescents and adults (Table 6 and 7). These studies have shown that prolonged administration of non-myelosuppressive agents such as asparaginase, vinca alkaloids and steroids is feasible and tolerated in a substantial portion of adults. Based on these studies, pediatric based regimens have now been widely accepted as the standard of care for younger adults with ALL.

Table 5: Outcomes of adolescents and young adults treated on pediatric protocols.

<table>
<thead>
<tr>
<th>Study</th>
<th>Number</th>
<th>Age Range</th>
<th>Complete Remission</th>
<th>Outcomes</th>
<th>Overall Survival</th>
</tr>
</thead>
<tbody>
<tr>
<td>LALA 94 vs. FRALLE 93</td>
<td>FRALLE - 77</td>
<td>15 - 20</td>
<td>FRALLE - 94%</td>
<td>FRALLE - 5 year OS - 78% ± 11%; 5 year DFS - 72%, 5 year EFS - 67%; 5-year RFS --&gt; 77%</td>
<td></td>
</tr>
<tr>
<td>LALA - 107</td>
<td></td>
<td></td>
<td>LALA - 83%</td>
<td>LALA - 5 year OS - 45%; 5 year DFS - 49%; 5 year EFS - 41%; 5-year RFS --&gt; 49%</td>
<td></td>
</tr>
<tr>
<td>DCOG vs. HOVON 91</td>
<td>DCOG - 47</td>
<td>DCOG - 15 - 18</td>
<td>DCOG - 98%</td>
<td>DCOG - 15 - 18 - 4% (5 years TRM)</td>
<td></td>
</tr>
<tr>
<td>HOVON - 44</td>
<td></td>
<td></td>
<td>HOVON - 15 - 18 - 91%</td>
<td>HOVON - 15-18 - 25% (5 year TRM)</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>HOVON - 19 - 20 - 90%</td>
<td>HOVON - 19 - 20 21% (5 year TRM)</td>
<td></td>
</tr>
<tr>
<td>UKALL vs ALL97 107</td>
<td>UKALL - 67</td>
<td>15-17</td>
<td>ALL97 - 98%</td>
<td>ALL97 - 5 year - 71%; 5 year EFS - 69%</td>
<td></td>
</tr>
<tr>
<td>Study</td>
<td>Number</td>
<td>Age Range</td>
<td>Complete Remission</td>
<td>Outcomes</td>
<td>Overall Survival</td>
</tr>
<tr>
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<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>UKALL</td>
<td>5 year EFS - 49%</td>
</tr>
<tr>
<td>ALL97 - 61</td>
<td></td>
<td></td>
<td>94%</td>
<td></td>
<td>56%</td>
</tr>
<tr>
<td>DFCI92</td>
<td>DFCI - 51</td>
<td>15 - 18</td>
<td></td>
<td></td>
<td>5 year OS - 81%</td>
</tr>
<tr>
<td>PETHEMA93</td>
<td>Age 15 - 18 = 35</td>
<td>15 - 30</td>
<td>Age 15 - 18 --&gt; 94%</td>
<td></td>
<td>6 years EFS - 60%</td>
</tr>
<tr>
<td></td>
<td>Age 18 - 30 = 46</td>
<td></td>
<td>Age 18 - 30 --&gt; 100%</td>
<td></td>
<td>6 years EFS - 63%</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>77%; 6 years EFS - 63%</td>
</tr>
<tr>
<td>CCG vs. CALGB25</td>
<td>CCG - 162</td>
<td>15 - 20</td>
<td>CCG - 90%</td>
<td></td>
<td>67%; 7 year EFS - 63%</td>
</tr>
<tr>
<td></td>
<td>CALGB - 124</td>
<td>16 - 20</td>
<td>CALGB - 90%</td>
<td></td>
<td>46%; 7 year EFS - 34%</td>
</tr>
<tr>
<td>CCG 196140</td>
<td>262</td>
<td>16 - 21</td>
<td></td>
<td></td>
<td>77.5%; 5-year EFS - 71.5%</td>
</tr>
<tr>
<td>St. Jude XV108</td>
<td>45 on XV study</td>
<td>15 - 18</td>
<td></td>
<td></td>
<td>87.9%; 5 years EFS - 86.4%</td>
</tr>
</tbody>
</table>

**Table 6: Outcomes of adult patients using paediatric protocols.**

<table>
<thead>
<tr>
<th>Study</th>
<th>Number</th>
<th>Age Range</th>
<th>Median</th>
<th>Complete Remission</th>
<th>Outcomes</th>
<th>Overall Survival</th>
</tr>
</thead>
<tbody>
<tr>
<td>GRAALL vs. LALA - 9494</td>
<td>GRAALL - 225</td>
<td>15 - 55</td>
<td>31</td>
<td>GRAALL - 93.5%</td>
<td>GRAALL: OS 3.5 years --&gt; 61%; EFS - 57%; CIR - 31%</td>
<td></td>
</tr>
<tr>
<td></td>
<td>LALA - 712</td>
<td>15 - 55</td>
<td>29</td>
<td>LALA - 88%</td>
<td>LALA: OS 3.5 years --&gt; 41%; EFS - 33%; CIR - 55%</td>
<td></td>
</tr>
<tr>
<td>DFCI20</td>
<td>DFCI - 85</td>
<td>18 - 60</td>
<td>37</td>
<td>&lt; 35 = 98%</td>
<td>0% age &lt; 35</td>
<td>3 year OS - 67%</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>&gt; 35 yo -&gt; 81%</td>
<td>8% age - 36-49</td>
<td>5 Year OS - 63%</td>
</tr>
</tbody>
</table>

Last revision October 2023
<table>
<thead>
<tr>
<th>Study</th>
<th>Number</th>
<th>Age Range</th>
<th>Median</th>
<th>Complete Remission</th>
<th>Mortality</th>
<th>Overall Survival</th>
</tr>
</thead>
<tbody>
<tr>
<td>DFCI45</td>
<td>Age 17 - 60 --&gt; 156</td>
<td>17 - 60 37</td>
<td>17 - 34 = 99%</td>
<td>73% age &gt;50</td>
<td>DFCI Age 18 - 35: 3 year OS - 83%; RFS - 77%</td>
<td>Combine d - 89%</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>34 - 50 = 87%</td>
<td></td>
<td>DFCI Age 35 - 60: 3 year OS - 52%; RFS - 60%</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>50 - 60 = 90%</td>
<td></td>
<td>DFCI Age 18 - 35: 5 year OS - 80%</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Combine d - 93%</td>
<td></td>
<td>DFCI Age 35 - 60: 5 year OS - 50%</td>
<td></td>
</tr>
<tr>
<td>DFCI109</td>
<td>51</td>
<td>60 - 79 65</td>
<td>BCR - ABL Positive --&gt; 81%</td>
<td>20%</td>
<td>BCR - ABL Positive --&gt; 47.3 % at 5 years</td>
<td>BCR - ABL Negative --&gt; 40.5% at 5 years</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(12&gt;70 )</td>
<td>BCR - ABL Negative --&gt; 71%</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>DFCI95</td>
<td>92</td>
<td>18-50 28</td>
<td>85%</td>
<td>1/92 patients</td>
<td>4 Year OS 67%; DFS - 69%; EFS - 58%</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>18-29 48</td>
<td>16/18 = 89% T-cell</td>
<td></td>
<td>Age 18 - 29: 4 Year OS - 68%; DFS - 70%; EFS - 55%</td>
<td></td>
</tr>
</tbody>
</table>
**Study Number | Age Range | Age Median | Complete Remission | Outcomes | Overall Survival**

<table>
<thead>
<tr>
<th><strong>Study</strong></th>
<th><strong>Number</strong></th>
<th><strong>Age</strong></th>
<th><strong>Complete Remission</strong></th>
<th><strong>Outcomes</strong></th>
<th><strong>Overall Survival</strong></th>
</tr>
</thead>
<tbody>
<tr>
<td>DFCI</td>
<td>30</td>
<td>51-72</td>
<td>44</td>
<td>62/74 = 84% B-cell</td>
<td>Age 30 - 50: 4 Year OS - 65%; DFS - 67%; EFS - 61%</td>
</tr>
<tr>
<td></td>
<td></td>
<td>58</td>
<td></td>
<td>14/18 = 78% Ph - +ve</td>
<td>T-cell ALL: 4 year OS - 76%; DFS - 87%; EFS - 77%</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>64/74 = 86% Ph - -ve</td>
<td>Ph -ve/B-cell: 4 Year OS - 68%; DFS - 66%; EFS - 57%</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Ph +ve/B-cell: 4 year OS - 53%; DFS - 54%; EFS - 42%</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>WBC &lt; 20: 4 year OS - 80</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>WBC &gt; 20: 4 year OS - 45%</td>
</tr>
<tr>
<td>CALGB</td>
<td>111</td>
<td>17-39</td>
<td>24</td>
<td>age &gt; 50 = 67%</td>
<td>62% at 1 year; &lt; 20% at 3 years (based on survival curve)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Age &gt; 50 = 10%</td>
<td>EFS 59% at 3 years OS 73% at 3 years</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>3%</td>
</tr>
</tbody>
</table>

**CNS Prophylaxis and Treatment:**

Because of the high risk (up to 50%) of CNS relapse, CNS prophylaxis is an essential part of any treatment regimen for ALL. Standard approaches include the use of repeated (11-12) doses of intrathecal (IT) chemotherapy. This has been demonstrated in many studies to reduce the risk of CNS relapse to 10% or less. Although many protocols use triple intrathecal chemotherapy with methotrexate, cytarabine and corticosteroids, a recent pediatric randomized study did not show any difference in CNS relapse rates between this approach and single-agent methotrexate.112 A baseline LP is required to rule out CSF disease at diagnosis. If positive, the usual approach is to administer intrathecal chemotherapy twice weekly until the CSF has been adequately cleared (as evidenced by 3 consecutive negative results). Although the standard cerebrospinal fluid (CSF) diagnostic approach consists of morphologic analysis of a cytospin preparation, studies have shown that flow cytometry of CSF provides greater sensitivity,113 and may thus be a more reliable indicator of blast clearance.

Most older treatment protocols incorporated prophylactic cranial radiation; however, this adds significant short-term and potential long-term toxicities. Recent pediatric studies have demonstrated that routine cranial radiation can be safely deleted without adversely impacting CNS relapse rates;114
this is particularly the case if high-dose intravenous methotrexate, which has good CNS penetration, is used. However, if evidence of fixed leptomeningeal disease is present based on the presence of focal neurologic symptoms and corresponding MRI findings, CNS radiotherapy is indicated as intrathecal chemotherapy alone is usually insufficient.

**Provincial Recommendations for Treatment of BCR-ABL negative ALL**

**Adolescents and Young Adults (AYA):**
Adolescents and young adults have been variably defined as those aged 15 – 35 (40) years old. A multitude of accumulating evidence suggests that this group of patients is best treated with a pediatric or a pediatric inspired protocol. Ram et al. conducted a systematic review of 11 studies that compared adult and pediatric protocols.115 Overall, all-cause mortality, relapse rates and non-relapse mortality were lower in the pediatric groups whereas CR rates and EFS were higher in the pediatric groups. It is important to note that in many of these studies the median age ranged from 12.9 – 31 with only two studies including patients over age 55.

Boissel and colleagues compared the outcomes of adolescents aged 15 – 20 years of age treated with the adult LALA-94 vs. the pediatric FRALLE-93 protocol 90. The LALA-94 trial included 100 patients (median age 17.9 years) and the FRALLE-93 trial included 77 patients (median 15.9 years) 15- 20 years of age. The CR rate was significantly higher in the FRALLE-93 trial (94 %) than the LALA-94 trial (83 %). The 5 year-EFS was also worse in the LALA-94 trial (67 %) compared to 41 % in the FRALLE-93 trial. In the Dutch study by de Bont and colleagues 91, adolescents aged 10 – 20 years old were treated with the Dutch Childhood Oncology Group (DCOG) or adult Dutch-Belgian Hemato-Oncology Cooperative Study Group (HOVON) adult protocol. For 15–18-year-old adolescents, the 5- year event-free survival (EFS) when treated on pediatric protocols was 69%, which is significantly higher than when treated on adult protocols (34%).

Stock and colleagues25 assessed the outcomes of 321 adolescents and young adults ages 16 – 20 years treated on either the paediatric Children’s Cancer Group (CCG) protocols (N= 197, median age 16) or the CALGB group protocols ( N= 124, median age 19). EFS at 7 years for the 25 CALGB patients aged 16 to 17 years was 55% which was not different that the CCG patients (P =.49). Notably, 18-20 year olds treated with the CALGB protocol fared significantly worse compared to 18-20 year olds treated with the CCG protocol (7 year EFS 29% vs. 57%). More frequent and earlier CNS prophylaxis was incorporated into the CCG trials leading to a lower CNS relapse rate of 1.4% in the CCG arm and 11 % in the CALGB arm.

In the most recent study the GRAALL study group evaluated the outcomes of 225 adult patients aged 15 to 60 treated with the GRAALL-2003 protocol and compared the outcomes with 712 patients aged 15 – 55 treated on the adult LALA-94 protocol. The overall CR rate for the 225 GRAAL-2003 treated patients was 93.5% with 6% induction mortality. A significantly longer OS was observed in patients aged 15 - 45 at 42 months when treated with the GRAALL-2003 protocol. Storring and colleagues
also assessed the outcomes of adults treated with the modified DFCI paediatric regimen.\textsuperscript{20} Between 2000 and 2006 85 adults aged 18 -60 (median age 37) with BCR-ABL negative ALL were treated. CR was achieved in 89% of cases. Of these, only age predicted for achievement of CR. For patients aged ≤ 35 years, the complete response rate was 98% (41/42). For patients age < 35 the median 3 year RFS was 77% and the median 3 year OS was 83%.

**Recommendations for AYA patients:**
Notwithstanding the potential barriers to administration of a paediatric protocol, we recommend that all adolescents and young adults patients be treated with a paediatric protocol. In Alberta, as in many other Canadian centers, the standard regimen is the modified Dana Farber Cancer Institute (DFCI) protocol. The original DFCI protocol is shown in Appendix A. The modified AL.4 protocol used in National Cancer Institute of Canada ALL study is shown in Appendix B. The modified protocol as used by the Princess Margaret Hospital (PMH) is shown in Appendix C. Patients with co-morbidities, felt to be ineligible for the full protocol, may require modifications to minimize toxicity.

The native E. coli asparaginase used in the DFCI 91-01 protocols is no longer available in Canada, and has been replaced by pegylated asparaginase (Pegasparagase). This should be given no more frequently than once every 3 weeks in adults due to its long half-life (see Appendix C and D).

**Patients Aged 35(40)-60:**
The use of pediatric protocols for the treatment of adolescents has inspired some groups to explore the use of pediatric protocols in older adults however the data are not clear whether this represents a significant difference relative to adult protocols. When using the paediatric GRAALL-2003 protocol improvements in EFS and OS were noted at 42 months but only for patients < age 45.\textsuperscript{94} Similarly, Storrting and colleagues, when using a modified DFCI paediatric regimen\textsuperscript{20} for adults aged 18 -60 noted that the complete response rate decreased with increasing age – from 86% for patients aged 36–49 years, to 73% for patients aged ≥ 50 years. In addition, there was a trend for an increase in induction mortality by age: 0% for age ≤ 35 years, 8% for age 36–49 years, and 20% for age ≥ 50 years. In addition, age > 35 was significantly associated with inferior 3 year - OS (83% vs. 52%). In an updated analysis,

Brandwein et al. assessed the outcomes of 156 adults with BCR-ABL negative ALL treated with a modified DFCI. Again, the CR rate decreased with increasing age. The CR rate amongst those age <34 was 99% while for those age 34-50 the CR rate was 87% and those aged 50-60 was 90%. The 5 year OS amongst patients aged <34 was 80% while those for patients aged 34-50 was 50% and those aged 50-60 was 62%. Whether the decreased rate of survival seen in patients aged 34-50 was due to an increased use of allo- SCT in this age group is unknown. DeAngelo and colleagues recently presented the results of their studies on the use of the DFCI protocol for patients 18-50 years old. They found no statistically significant difference in the outcomes for patients that were 18-29
years old or those that were 30-50 years old in terms of 4 years DFS (70 vs. 67%), 4 year EFS (55 vs. 61%) or 4 year OS (68 vs. 65%).

**Recommendations for Patients Aged 35(40)-60:**
These data suggest that although results in patients 30-60 years old may be inferior to those in adolescents and young adults a use of a pediatric protocol may benefit these patients with 3-5 year OS improving from an historic 35-40% to 50-70%. The lower OS observed in some studies may be related to the preferential use of allo-SCT in this cohort of patients. Nonetheless, we recommend that patients in this age range continue to be treated with the DFCI protocol (Appendix A-C). However, given the increased toxicities seen in middle-aged adults, some patients, particularly those age 50-60, may require changing to a less toxic regimen such as the modified DFCI protocol for older patients (see below).

**Patients Aged >60:**
*Fit patients:* The outcome of patients above age 60 treated with standard ALL chemotherapy has been generally poor. Larson et al. using the CALGB 8811 protocol noted a 3 year OS of only 17% with a median survival of only 1 month. Brandwein et al. noted similar results when using a multitude of different adult based chemotherapy regimens. In their study the 3 year OS was only 18.4%. Using the HyperCVAD protocol Kantarjian et al. noted a 5 year OS of 17%. Thus, it appears that the majority of older patients with ALL will succumb to their disease although some older patients may remain long-term survivors.

Whether a paediatric inspired regimen might benefit this older subset of patients was investigated by Martell et al. who evaluated the outcomes of 51 patients treated with a modified DFCI paediatric regimen at the PMH (Appendix B). Modifications from the full-dose DFCI 91-01 protocol during induction included the substitution of dexamethasone for two 4-d pulses instead of daily prednisone, reduction of the methotrexate dose from 4 g/m2 to 40 mg/m2, reduction in the asparaginase dose from 25 000 to 12 000 iu/m2 and removal of one vincristine dose. In the CNS prophylaxis phase cranial radiation was removed. In the intensification phase seven cycles were given instead of 10, the dexamethasone dose was reduced to 6 mg PO BID and the asparaginase reduced to 6000 iu/m2 from 12500 iu/m2. In the maintenance phase parenteral methotrexate was switched to oral, and the dexamethasone dose was again reduced from 6 mg/m2 BID to 6 mg PO BID. For patients who developed progressive grade ≥2 neuropathy, intravenous vinblastine 10 mg was substituted for vincristine. Median age was 65 (60 – 79), 12 patients were over the age of 70, 35 patients were BCR-ABL negative and 16 were BCR – ABL positive. The CR rate was 75% for the entire cohort with 20% induction mortality. The estimated 5-year OS was 40.5% for the BCR-ABL1 negative patients, and 47.3% for the BCR-ABL1+ patients but there was no significant difference in OS between these two groups. The estimated 5-year OS for BCR-ABL1 negative patients presenting with low WBC (defined as <30 x 10⁹/L for B-ALL or <100 x 10⁹/L for T-ALL) was 44.3% respectively.
Unfit patients: Many elderly patients are unfit for such intensive chemotherapy approaches, due to comorbidities and overall frailty. For those patients, palliative low-dose chemotherapy may be used. The Spanish PETHEMA group recently reported outcomes in such patients using weekly vincristine plus dexamethasone with IT chemo, followed by maintenance with 6MP and methotrexate\textsuperscript{157}. The regimen was well-tolerated; a 54% CR rate was obtained with 10% early deaths; the median OS was 7.6 months and one year OS approx. 20\% \textsuperscript{157}. In some patients with better ECOG PS a single dose of pegasparagase 500 U/m\textsuperscript{2} may be added to induction, but increases the risk of liver toxicity. If a CR is achieved and PS remains good, a slightly more intensified maintenance chemotherapy using the POMP regimen (vincristine, prednisone, 6-MP and weekly oral methotrexate) may be used in an attempt to prolong remission duration. If neurotoxicity becomes an issue, the vinblastine may be used, or the vinca can be deleted.

**Recommendations for Patients Aged >60:**
Given that some medically fit patients above age 60 may be cured of ALL we recommend that eligible patients over age 60 be treated with curative intent therapy. Although, there are no randomized studies, based upon data from the PMH group, the modified DFCI provided superior results relative to historical controls. We therefore recommend that the modified DFCI protocol (Appendix C) or similar regimen be used for older ALL patients. A more recent report from the Spanish PETHEMA group also showed reasonably good activity with their protocols, although relapse rates remained high.\textsuperscript{117}

Patients over age 75, or those age 60-75 with major co-morbidities precluding intensive chemotherapy, should be considered for palliative chemotherapy with corticosteroids and vincristine followed by maintenance therapy. Such patients should continue to receive central nervous system prophylaxis and may benefit from incorporation of a CNS phase. Patients not tolerating DFCI-type intensification, or judged to be unfit for such treatment, may be moved directly to the DFCI maintenance phase.

**Philadelphia Chromosome (BCR-ABL) Positive ALL:**
Tyrosine kinase inhibitors (TKI’s) have become the standard of care for Ph+/BCR-ABL+ ALL and have led to improvements in the outcomes for all age groups.\textsuperscript{1} The earliest TKI used in ALL was imatinib. Although, different chemotherapy regimens and schedules of imatinib have been assessed, all showed improvements in overall survival and reduction in the relapse rate.\textsuperscript{118-120} In the first large study done by the French GRAAPH-2003 group, the combination of imatinib with induction and postremission therapy\textsuperscript{121} led to higher estimated OS (65% vs. 39%), lower CIR (30% vs. 49%) and improved DFS (51% vs. 31%) at 18 months in comparison to historical controls treated with the LALA-94 protocol. An Italian study also reported 5-year OS of 38\% and DFS of 39\% with imatinib combined with chemotherapy and again these results were significantly superior to a historical cohort receiving chemotherapy alone.\textsuperscript{122} In the large UKALL/ECOG2993 trial investigators reported an improved post-induction CR rate with imatinib, improved 4 year OS, EFS and a considerable reduction in relapse risk in the imatinib cohort.\textsuperscript{123} Lastly, The M.D. Anderson group also combined
imatinib with their standard hyper-CVAD protocol and reported CR rates of 93% as well as results that were substantially superior to their retrospective results with chemotherapy.124

Second and third generation TKI’s have also been assessed in Ph+ ALL.125-128 Kim et al evaluated the outcomes of Nilotinib with multiagent chemotherapy for adult patients with newly diagnosed Ph+ve ALL. After achieving CR, subjects received either 5 courses of consolidation, followed by 2-year maintenance with nilotinib, or allo-HCT. Amongst the 90 evaluable subjects the CR rate was 91%, the 2-year RFS was 72% and the 2-year OS was 72%. Unlike nilotinib, dasatinib has the ability to penetrate the CNS. Single agent dasatinib is associated with short-term cytogenetic remission and median relapse free survivals of only 3.3 months. Studies of dasatinib in combination with chemotherapy, however, have been associated with excellent response rates approaching 100% with minimal toxicity.72,125 Ponatinib was evaluated in Ph+/BCR-Abl+ by Jabbour and colleagues for patients up to age 60 with previously untreated Ph +ve ALL. Ponatinib 45 mg was given daily for the first 14 days of cycle 1 then continuously each subsequent cycle of hyper-CVAD. Patients in complete remission received maintenance with ponatinib 45 mg daily with vincristine and prednisone monthly for 2 years followed by ponatinib indefinitely. The 2-year EFS rate was 81% (95% CI 64-90) for the 37 patients enrolled in the study.128

Several investigators have combined TKI’s with paediatric based regimens used for the treatment of adult patients. Thyagu et al. evaluated the outcomes of 32 ALL patients age 18-60 with Philadelphia positive ALL treated between 2001 and December 2008 with imatinib. Ninety-four percent of patients (94%) achieved a CR and, of the 28 patients proceeding to intensification therapy, 16 received a HSCT. The 3-year OS was 56% in the transplanted group vs. 50% in the non-transplanted group.129 They also noted increased rates of peripheral neuropathy, ileus, myopathies, deconditioning, infections and abnormal liver enzymes in a high proportion of patients. Therefore, a number of modifications were made to the original DFCI protocol.129 Using an unmodified DFCI protocol, Deangelo et al. reported similar results amongst 18 Ph +ve patients treated with imatinib, 11 of whom went on to transplant. The 4 year DFS was 54% and 4 year OS was 53%.95 Martell et al. also treated 16 BCR-ABL positive patients aged 62 – 75 with a modified DFCI protocol together with imatinib. The CR rate was 81% with an induction mortality of 19% and the estimated 5-year OS was 47.3% for the BCR-ABL1+ patients.109

Monotherapy with TKI has been explored in several studies particularly in elderly patients, who had an extremely poor outcome with chemotherapy alone. Ottmann et al evaluated imatinib monotherapy in 27 patients older than 55 years of age, and observed CR in 26 patients and a partial remission in the remaining patient.130 Vignetti et al. treated patients aged 61 to 83 years with Ph–+ ALL with a 45-day induction of imatinib 800 mg daily plus prednisone (40 mg/m² daily). Post-remission therapy was not specified. All 29 assessable patients (100%) experienced a CR. At 12 months, the OS and DFS probabilities were 74% and 48%, respectively.131
The French GRAALL group conducted a large randomised controlled trial comparing a reduced intensity induction with imatinib, vincristine and dexamethasone (Arm A) to standard imatinib/hyper-CVAD A part (Arm B) in 268 adults with Ph+ve ALL up to age 60; both arms then received a second cycle with Hyper-CVAD B part. The CR rate was higher in arm A than in arm B (98% vs 91%), mainly related to a lower induction death rate in Arm A, whereas the MMolR rate was similar in both arms (66% vs 64%). The OS was superior in the patients that subsequently proceeded to allogeneic HSCT; however, on subgroup analysis, patients who had achieved MRD negativity (defined as a >4 log reduction in BCR-ABL transcripts by PCR) by the end of the second induction cycle had a similar RFS with or without allogeneic HSCT. In contract patients who were MRD positive at that timepoint had a significantly better RFS with allogeneic HSCT, compared to those who were not transplanted. Therefore, MRD assessment with this regimen could be used to help identify a high-risk cohort for whom HSCT was beneficial.

Foà et al. evaluated a similar regimen consisting of dasatinib (70 mg twice daily for 84 days) together with prednisone 60 mg/m² daily for 32 days plus two doses of intrathecal methotrexate in untreated patients with Ph+ve ALL. Post remission therapy was not defined with two patients receiving no further therapy, 19 continuing on TKI only (16 dasatinib, 2 imatinib and 1 imatinib-dasatinib), 10 receiving intensive chemotherapy with TKI, 4 having an autograft and 18 receiving an allogeneic HSCT. Overall, 53 patients aged 24 – 76 were treated. A CR rate of 100% was seen with 92.5% by day 22 and no deaths occurred during induction. Of the patients that had MRD measured by PCR 10 achieved levels lower than 10⁻³ and 8 had a complete molecular remission. Twenty-three patients relapsed after completing induction with 12/17 relapsing patients showing a T315I mutation. This trial demonstrated impressive remission rates for dasatinib and steroids only but at the same time underscored the importance of adding intensive chemotherapy and/or HSCT to maintain durable remissions.

More recent studies suggest that the treatment landscape is changing. The MD Anderson group evaluated the use of ponatinib + Hyper-CVAD chemotherapy in fit patients with BCR-ABL+ ALL. The 3-year EFS of 69% and OS of 84%, without transplant, were significantly superior to a historical group that had received dasatinib + Hyper-CVAD. These results suggest that using ponatinib with intensive chemotherapy upfront may potentially circumvent the need for transplant in CR1 for most patients. More recently the Italian GIMEMA group reported on a chemo-free regimen using dasatinib + blinatumomab. The CR rate was 98%, with 70-80% eventually achieving a complete molecular response; at a median follow-up of 18 months, the OS was 95% and DFS 88%. The MD Anderson group is currently using a combination of ponatinib + blinatumomab for 4 cycles, followed by ponatinib maintenance therapy, as their standard frontline regimen; the CR rate was 100% and 86% achieved a complete molecular response. While very promising, follow-up is brief, and these treatments are not currently approved as upfront therapy for BCR-ABL+ ALL.
Recommendations Philadelphia Chromosome or BCR-ABL Positive ALL:
All patients with Ph+/BCR-ABL+ve ALL should receive a TKI combined with chemotherapy. No randomized studies are available comparing different TKI’s in adults; therefore, a firm recommendation regarding the choice of TKI upfront cannot be made. Currently, only imatinib (600-800 mg daily) is approved as first line therapy. Patients intolerant to imatinib should be switched to another TKI such as dasatinib; dasatinib may also be preferred for patients with CNS disease due to its excellent CNS penetration. Patients with documented T315I BCR-ABL mutations, or those refractory to or progressing on, a 2nd generation TKI, should be treated with ponatinib.

Fit BCR-ABL+ve ALL patients should be treated with a TKI combined with induction and post-remission chemotherapy. Given recent studies showing higher remission rates and decreased mortality using corticosteroids, TKI + vincristine during induction, this combination is recommended as initial induction therapy (see Appendix F); however, such patients require additional intensive chemotherapy +/- HSCT to maintain a durable remission.

Given data suggesting increased toxicity of asparaginase and vincristine with TKI, asparaginase should be deleted, and vinblastine substituted for vincristine, in the post-remission phases. Patients above age 65, particularly, those with major co-morbidities, poor performance status or ineligible for SCT should be treated with steroid and TKI +/- vincristine or vinblastine. Post induction therapy should be individualized based upon patient tolerance. Patients not transplanted should continue TKI indefinitely.

BCR-ABL monitoring by quantitative RT-PCR should be performed in all patients, post-induction and then every 3 months. Patients with inadequate response or loss of response should be switched to a second or third generation TKI as indicated and, depending on circumstances, may require switching from chemotherapy to blinatumomab.

Role of Allogeneic Stem Cell Transplantation (SCT)

Ph-/BCR-ABL- ALL:
In the largest adult ALL trial to date (MRC-ECOG UKALLXII/E2993) patients in first complete remission < 50 years were assigned to alloSCT if they had a compatible sibling donor while others were randomized to consolidation/maintenance therapy for 2.5 years or to autologous transplant and no further therapy. Patients were considered to be high-risk if they had age >35, WBC >30,000/ mL for B-lineage or WBC >100,000/mL for T-ALL, Philadelphia chromosome positivity, t(4;11), t(8;14), complex karyotype, low hypodiploidy or triploidy. All other patients were considered standard risk. In a donor versus no-donor analysis, patients with a sibling donor had improved OS than those with no donor (53% vs. 45%). In the standard risk group, the OS at 5 years was 62% for patients with a donor compared to 52% for patients with no donor (P=.02). In contrast, for high-risk patients OS was 41% for those with a donor vs. 35% for those without a donor (p=0.2) likely due to the high non-relapse
mortality. The 10 year relapse rate was substantially lower in patients with a donor (24% for standard risk and 37% for high-risk) versus those without a donor (49% standard risk and 37% high-risk). The Spanish PETHEMA group reported similar findings in their cohort of high-risk patients defined as those with age 30-50 years old, WBC count >25,000/mL, Ph+ve, t(4;11)/other 11q23 rearrangements, or t(1;19). In contrast, two French studies both reported that there was an advantage for high-risk patients having a donor.

A meta-analysis of 7 studies that included 1274 patients also noted improvements in overall survival for patients with donors relative to those in the no-donor groups undergoing allogeneic stem-cell transplantation. When only high-risk patients were analyzed, the survival advantage was greater. More recently, Gupta et al. conducted a meta-analysis using individual patient data from a total of 20 trials that included a donor no-donor comparison. Overall survival at 5 years was significantly better in the donor arm (49.9% vs 42.7%, p=.003). However, because TRM was much higher in those age >35 both in the donor and no-donor arms there was no difference in OS in this age group. Interestingly, unlike in previous studies and using standardized definitions of high/standard risk, there was no evidence that survival differed by risk category. It was therefore concluded that patients age<35 benefit from an allo-SCT regardless of risk group. Furthermore, by reducing TRM, RIC has the potential to extend the benefit of allo-SCT to those age>35.

Despite the above data, it remains unclear whether adult patients treated with paediatric protocols would gain a benefit from SCT. In their study of a 156 BCR-ABL–ve patients treated with the DFCI protocol the 5 year OS amongst patients receiving an allogeneic SCT was 44% while for those not undergoing a SCT the survival was 74% with the difference possibly related to transplant related mortality. Seftel and colleagues compared 422 Ph-ve ALL patients aged 18-50 years with 108 patients receiving DFCI chemotherapy. Expectedly, treatment related mortality was higher in those receiving a SCT (37% vs. 6%). At 4 years, the incidence of relapse was similar for those receiving SCT and chemotherapy (24% vs. 23%), however, both DFS and OS were improved for those receiving chemotherapy alone (40% vs. 71% for DFS and 45% vs. 73% for OS). Dhedin and colleagues further assessed the role of allogeneic stem cell transplantation in Ph-ve ALL adult patients with at least 1 conventional high-risk factor treated with the pediatric inspired GRAALL 2003 and 2005 protocols. In all, 522 patients age 15 to 55 years old were candidates for SCT in first complete remission. Among these, 282 (54%) received a transplant in first complete remission while 240 (46%) did not. As with previous studies, the lower CIR observed in the SCT was counterbalanced by a higher NRM. When analyzing SCT in CR1 as a time-dependent event RFS and OS were not significantly improved in the SCT cohort. No significant effect of SCT on RFS was noted in patients younger or older than age 45 or on any prespecified baseline risk factor. RFS and OS were significantly longer, however, in patients who presented with morphologic poor early BM blast clearance or in late CR patients. Furthermore, SCT was associated with longer RFS in patients with postinduction minimal residual disease (MRD) >10^-3 but not in good MRD responders.
For pediatric-based protocols, asparaginase is a critical agent in achieving the high cure rates reported. There is evidence in both pediatric and adult studies with the DFCI protocol that the inability to deliver the intended asparaginase dosing during intensification (defined as \( \geq 80\% \)) is associated with a higher relapse rate.\(^{20,137}\) Therefore, the inability to deliver effective asparaginase dosing (e.g. due to pancreatitis) places the patients in a higher risk category and warrants consideration of allogeneic HSCT.

**Recommendations for Ph-/BCR-ABL- ALL:**
These data suggest that patients with Ph-/BCR-ABL -ve ALL treated with a paediatric protocol who attain an MRD negative complete remission are at low risk of subsequent relapse and will not benefit from an allogeneic SCT. It is therefore recommended that such patients not proceed with allogeneic SCT, unless they are unable to complete the protocol. Given that patients with MLL (KMT2A) rearrangements remain a high-risk group with pediatric based regimens, SCT may be a reasonable recommendation for younger ALL patients with this abnormality. Failure to achieve a complete hematologic remission after the first induction cycle is also generally considered a high risk features; transplantation is also recommended for these patients. Although high presenting WBC has been identified in the past as a high-risk feature, its prognostic value is superceded by MRD based on data from GRAALL. Failure to deliver effective asparaginase dosing during intensification should also warrant consideration of allogeneic SCT.

Patients who are MRD +ve at \( >10^{-3} \) (or >0.1%) after induction therapy, or \( >10^{-4} \) (<0.01%) at 16-18 weeks, are at higher risk of relapse and may benefit from transplant. Such patients should be considered for allogeneic HSCT, optimally after receiving further cytoreduction to attain an MRD negative state prior to transplant.

**Role of SCT in Patients with Ph/BCR-ABL Positive ALL:**
The UKALLXII/ECOG 2993 study evaluated the outcome of allo-SCT in Ph+/BCR-ABL+ patients younger than 55 years of age achieving complete remission with an adult ALL protocol. Of the 267 patients, 76 (28%) proceeded to alloHSCT in first CR (45 with cells from a sibling and 31 with cells from a MUD) whereas 86 received chemotherapy alone. The median EFS for all 267 Ph+ patients was 9 months and the median OS was 13 months. At 10 years, OS was 39% for sib alloHSCT, 31% for MUD alloHSCT, and 13% for chemotherapy. Comparing the outcome after any alloHSCT with the outcome after chemotherapy alone, OS, EFS, and RFS were all significantly superior for patients receiving any alloHSCT over those receiving chemotherapy alone. Whereas the leading cause of death in chemotherapy treated patients was relapse, the leading cause of death after transplantation was TRM, which was 27% after sib HSCT and 39% after MUD HSCT.\(^{31}\)

Limited information is available comparing adult Ph+ patients receiving a paediatric protocol with those undergoing an allo SCT. Thyagu et al. treated 32 patients with Ph+ ALL with DFCI protocol together with imatinib. Of the 28 patients proceeding to intensification therapy, 16 underwent an allo-
SCT. The 3 year OS was 56% in the transplanted group and 50% in the non-transplanted group.\textsuperscript{129} Recent studies in the pediatric setting noted that children receiving intensive multidrug chemotherapy with imatinib had a survival higher than 80% compared to 35% in historical controls, and even better than related or unrelated HSCT (>60%). In the recent US Children’s’ Oncology Group study, the outcomes at 3 years were not significantly different for those treated with chemotherapy plus imatinib compared with those assigned to alloHSCT.\textsuperscript{138} However, limited patient numbers precluded rigorous subgroup analysis.

Recent studies in adults are also demonstrating the impact of molecular status on relapse risk in Ph+ ALL. The MD Anderson Cancer Center\textsuperscript{139} found that molecular positivity at a variety of time points was associated with a significantly higher cumulative incidence of relapse. The French GRAAPH study\textsuperscript{132} found that the benefit of alloHSCT in CR1 was restricted to patients who were molecularly positive post-induction chemotherapy. On subgroup analysis, patients who had achieved MRD negativity (defined as a >4 log reduction in BCR-ABL transcripts by PCR) by the end of the second induction cycle had a similar RFS with or without alloHSCT. In contract patients who were MRD positive at that timepoint had a significantly better RFS with allogeneic HSCT, compared to those who were not transplanted. Therefore, MRD assessment with this regimen could be used to help identify a high risk cohort for which HSCT was clearly indicated.

In contrast, a study using nilotinib plus chemotherapy\textsuperscript{140} found that overall survival was superior in patients undergoing HSCT regardless of MRD status; however, 60% of patients achieving molecular negativity remained free of relapse at 30 months without a transplant. Taken together, these data suggest that some patients who achieve MRD negativity early on may remain relapse-free with continuing chemotherapy + TKI, potentially sparing them the toxicity associated with transplant.

**Recommendations for the Role of HSCT in Patients with Ph/BCR-ABL Positive ALL:**

Although allogeneic HSCT remains the standard post-remission approach for many patients with BCR-ABL positive ALL, patients who achieve early MRD negativity by PCR (e.g. after induction with the Chalandon protocol) may be continued on post-induction chemotherapy + TKI without a transplant; these patients should continue with indefinite TKI and regular PCR monitoring. However, all patients with persistent molecular positivity should be referred for allogeneic HSCT if otherwise eligible. Consideration should be given to switching to a more potent TKI such as dasatinib or ponatinib in these MRD+ patients prior to transplant. Furthermore, patients with subsequent recurrence of MRD detectable disease by PCR should also be referred for transplant.

**Role of Cranial Radiation**

CNS involvement at the time of presentation is uncommon in adults with ALL being reported in 5% to 7% of patients.\textsuperscript{141} Before the use of central nervous system (CNS) prophylaxis, the CNS was the most frequently reported site of initial recurrence in children with ALL, accounting for up to 75% of cases.\textsuperscript{141} Similarly, amongst adults with ALL, CNS recurrence occurs in approximately 30% of those in a hematological remission.\textsuperscript{1} Aur et al. published the results of St. Jude Total Study V in 1971.
demonstrating that 2,400 cGy cranial radiation and five doses of intrathecal methotrexate greatly diminished CNS relapse resulting in the first cures of childhood ALL.\textsuperscript{142} Subsequently, widespread incorporation of CNS prophylaxis led to the largest single “step up” in 10-year survival from approximately 20% to 60% among those diagnosed from 1970 to 1972 versus 1972 to 1975.\textsuperscript{142} Given the significant risk of CNS relapse current adult and pediatric protocols incorporate CNS prophylaxis with both systemic and intrathecal chemotherapy and/or radiation.

Cranial irradiation is an effective form of CNS-directed treatment, but its effectiveness is offset by substantial rates of secondary neoplasms, endocrinopathies, growth impairment, neurocognitive dysfunction, and neurotoxic effects. Amongst the pediatric population, Pui et al. found that prior cranial radiation was associated with a 20.9% cumulative risk of second neoplasms at 20 years in addition to a higher mortality and a greater likelihood of being unemployed when compared to an age matched general population.\textsuperscript{142} Subsequently, multiple trials compared CNS prophylaxis using intrathecal chemotherapy and/or intravenous methotrexate with cranial radiation. These trials demonstrated the efficacy of IT/IV therapy without cranial radiation leading to use of cranial irradiation for patients at especially high risk of CNS relapse and elimination of cranial radiation for infants or very young children, irrespective of their presenting features Jeha, 2009 #199. More recently, in the St Jude Total XV and Dutch Childhood Oncology Group acute lymphoblastic leukaemia-9 protocols cranial irradiation is replaced by triple intrathecal chemotherapy with methotrexate, hydrocortisone, and cytarabine for all newly diagnosed patients. The 5 year cumulative risk of isolated CNS relapse was 2.7% with the St Jude and 2.6% with the Dutch Childhood Oncology Group protocol similar to the relapse rates observed with prophylactic cranial irradiation (1.5–4.5%). In the largest study to date Vora and colleagues\textsuperscript{143} obtained data on 16623 patients aged 1 to 18 years old treated between 1996 and 2007 by 10 cooperative groups. In their analysis cranial radiation was associated with a reduced risk of relapse but only in patients with overt CNS disease at time of presentation. In other patients there was no difference in CNS relapse between those that received and did not receive cranial radiation. These data suggest that, in the context of a pediatric protocol prophylactic cranial irradiation can be safely omitted in patients in the context of effective intrathecal and systemic chemotherapy.

**Recommendations for the Role of Cranial Radiation:**
Given the above data we recommend that cranial irradiation not be used for CNS prophylaxis in patients receiving a pediatric protocol such as the DFCI if intrathecal or systemic chemotherapy is used, unless there is evidence of fixed intracranial disease at presentation on MRI.

**Supportive Care**
Supportive care remains an important aspect of the care of the ALL patient throughout all phases of treatment. All patients should be transfused packed red blood cells as per institutional guidelines and particularly for symptomatic anemia. Similarly, platelets should be transfused to maintain platelet counts > 10 x 10^9/L. All patients receive prophylaxis, and if indicated, treatment for tumour lysis
syndrome with allopurinol and/or rasburicase during induction therapy. Disseminated intravascular coagulation should be treated aggressively with clotting factor replacement. Patients with signs and symptoms of febrile neutropenia should be managed with broad spectrum antibiotics and intensive care support as necessary. As corticosteroids may mask fevers in such patients, broad spectrum antibiotics should be instituted for clinical suspicion of sepsis (e.g. unexplained hypotension) even in the absence of fever. Antifungal prophylaxis against Candida is recommended during induction due to the risk of Candida septicemia.

Clinicians should be continuously aware of both the short- and long-term consequences of potential toxicities associated with specific agents used in ALL and use prevention, treatment or dose adjustment as necessary:

1. **Corticosteroids** may lead to hyperglycemia or gastrointestinal reflux/gastritis in the acute setting. Patients are also at risk for osteoporosis and osteonecrosis (avascular necrosis, AVN) with long-term use. Patients should be treated with gastric protective agents (e.g. PPI), calcium, vitamin D and bisphosphonates. Patients developing persistent pain in hips or other joints should have an MRI scan to exclude AVN.

2. **Asparaginase** has been associated with hypersensitivity reactions, hyperglycemia, coagulopathy, hepatotoxicity, and/or pancreatitis. Liver function tests should be performed regularly and patients should be monitored for clinical signs/symptoms of pancreatitis. Triglyceride levels should also be monitored as severe hypertriglyceridemia may be seen and could potentially increase the risk of pancreatitis. The development of pancreatitis requires discontinuation of asparaginase due to the high risk of recurrence. Grade 2 or higher liver toxicities warrant consideration of a dose reduction with subsequent cycles (see below for monitoring).

3. **Anticoagulation prophylaxis** with adjusted dose low-molecular weight heparin during the intensification phase of the DFCI protocol has been associated with a reduced risk of VTE\(^1\) and should be considered due to the high risk of VTE (25-30%\(^2\)); patients should be closely monitored for clinical signs of VTE. If a VTE is diagnosed, full-dose anticoagulation should be instituted, and the asparaginase can be resumed after several weeks.

4. **Hypersensitivity reactions** should be carefully assessed to exclude infusion type reactions. If unclear, a serum asparaginase level should be obtained 3-7 days later to confirm rapid clearance of drug.\(^1\) If confirmed based on clinical suspicion and lack of detectable asparaginase, the patient should be switched to Erwinia asparaginase which should be given 3x/week due to its short half-life.\(^1\)

5. **Therapeutic drug monitoring:** Silent inactivation of asparaginase, due to rapid clearance by antibodies, may also occur which could compromise treatment efficacy. Because of this, it is recommended that all patients have a trough serum asparaginase level drawn at the end of the
first cycle of intensification therapy (i.e. days 19-22), targeting a level > 1.0 U/L. A trough level should be repeated at cycle 6, or anytime if suspected based on surrogate markers (e.g. normal LFT’s and fibrinogen levels). If the trough level is < 0.1 U/L, an asparaginase level should also be repeated at day 3-7 post-dosing with the next cycle; if still <0.1 U/L, this would be consistent with silent inactivation and warrants a switch to Erwinia (van der Sluis et al 2016). Asparaginase levels may also be useful in de-escalating doses in cases of severe toxicity; however, standardized approaches are lacking. Any dose reduction should be followed by a repeat trough asparaginase level with the next cycle.

6. **Vincristine** can cause neuropathies including peripheral neuropathy and ileus. Careful attention to a bowel regimen to prevent severe constipation (e.g. PEG +/- sennosides) should be done. Patients with progressive peripheral neuropathy should be switched to vinblastine (6 mg/m² to a maximum of 10 mg/dose).

7. Other drugs: Doxorubicin may lead to cardiomyopathy while methotrexate and 6-mercaptopurine can lead to hepatotoxicity. Patients requiring tyrosine kinase inhibitors may encounter a variety of general and TKI specific side-effects.

8. **Antimicrobial prophylaxis**: Patients are severely immunocompromised and are at high risk of serious infections, including neutropenic sepsis, *pneumocystis jiroveci* pneumonia (PJP), viral infections (HSV or VZV reactivation, severe outcomes with influenza or COVID-19 infection), oropharyngeal Candidiasis, etc. Filgrastim should be used as needed for severe neutropenia during intensification phases. PJP prophylaxis (SMX-TMP, dapsone or inhaled pentamidine) and valacyclovir should be used. Patient should be strongly encouraged to undergo vaccination against influenza and COVID-19, and confirmed infection should be treated in its early stages with appropriate antiviral therapy to prevent severe outcomes.

**Follow-Up**

For patients with ALL the highest risk of relapse remains within the first two years following completion of chemotherapy. Patients should have regular blood work for the first 2-3 years and every three months thereafter until 5 years. Patients with concerning laboratory features or clinical signs and symptoms for ALL relapse should be evaluated with repeat bone marrow studies.

**Treatment of Relapsed ALL**

Adult ALL patients experiencing hematologic relapse have a poor prognosis with conventional chemotherapy salvage regimens. CR2 rates have been in the 30-50% range. Second remissions are invariably brief unless followed by allogeneic HSCT, and OS is in the 10-20% range.147, 148 Recent studies have demonstrated the superiority of antibody-based therapies for relapsed patients with B-ALL. **Blinatumomab**, a BiTE antibody construct described earlier (see MRD Section), was
evaluated in the Phase III randomized TOWER trial \(^{156}\). Adult B-ALL patients with relapsed (CR1 < 12 months or ≥ CR2) or refractory disease were randomized to receive either blinatumomab for 2 cycles, followed by up to 3 consolidation cycles, or conventional salvage chemotherapy. The CR rate was higher with blinatumomab (44% vs. 25%, \(p=0.001\)), and OS was significantly longer in the blinatumomab arm (7.7 months vs. 4 months, \(p=0.01\)). Responses in the blinatumomab arm were influenced by tumour burden: The CR rate was 65% in those with < 50% bone marrow blasts, vs. 34% in those with ≥ 50% BM blasts; nevertheless, CR rates were higher in both subgroups as compared with the chemotherapy arm. Although this study only included patients with BCR-ABL negative ALL, a subsequent study demonstrated a 36% CR rate in patients with relapsed/refractory BCR-ABL+ B-ALL, all of whom had failed second or later generation TKI’s.\(^{149}\)

Inotuzumab ozogamicin is an anti-CD22 antibody conjugated to the toxin calicheamycin. The immunoconjugate is internalized into CD22+ B-cells, followed by release of the calicheamycin, which induces DNA strand breaks. The INO-VATE trial was a Phase III randomized study in patients with relapsed/refractory CD22+ B-ALL.\(^{150}\) Patients were randomized to receive inotuzumab or conventional salvage chemotherapy. The CR rate was significantly higher in the inotuzumab arm (81% vs. 29%, \(p<0.001\)); furthermore, nearly all CR’s in the inotuzumab group were MRD negative. The PFS in the inotuzumab group was also significantly longer (5 mos vs. 1.8 mos, \(p<0.001\)), and the OS was also longer (7.7 mos vs. 6.7 mos, \(p=0.04\)). CR rates with inotuzumab were not influenced as much by tumour burden (86.7% for those with <50% BM blasts, vs. 78% for those with ≥50% BM blasts). Notably, this study included patients with BCR-ABL+ ALL, where a 79% CR rate was reported.

In addition to being more effective, both antibody drugs were well tolerated. The major toxicities of blinatumomab included cytokine release syndrome and neurologic toxicity, both of which were usually grade 1-2 and treated successfully with corticosteroids. The major toxicities of inotuzumab were nausea, febrile neutropenia and veno-occlusive disease (VOD/SOS) of the liver. The latter was a particular issue in patients who received >3 cycles and in those who subsequently proceeded to allogeneic HSCT using a duel alkylator conditioning regimen.

For patients with relapsed/refractory T-ALL, there are currently no available antibody-based therapies outside of clinical trials. Conventional salvage chemotherapy, using a non-cross resistant regimen (e.g. Hyper-CVAD), may be used, or nelarabine. The latter is a prodrug of ara-G which is converted to ara-GTP, after which it is then incorporated into DNA, resulting in cytotoxicity. It has demonstrated single agent activity in relapsed/refractory T-ALL, with a 31% CR rate and 41% ORR.\(^{151}\) However, responses are not durable, and require subsequent allogeneic HSCT if treated with creative intent. Major toxicities of nelarabine include myelosuppression and neurotoxicity. Neurotoxic effects may be either central or peripheral, and may be severe and irreversible.
Recommendations for Relapsed ALL:
Based on the two pivotal TOWER and INO-VATE trials, patients with relapsed or refractory BCR-ABL negative B-ALL should be treated with either blinatumomab or inotuzumab as salvage therapy. Although there are no clear recommendations with respect to the choice of agent, inotuzumab would generally be preferred in patients with higher disease burdens (> 50% BM blasts), based on higher responses rates in this group. In patients with lower disease burden, blinatumomab with be preferred for patients who are intended to proceed to subsequent allogeneic HSCT, due to the risk of VOD/SOS with inotuzumab.

For patient who receive inotuzumab with an intention to proceed to HSCT, patients should optimally not receive more than 2 cycles, and dual alkylator HSCT conditioning regimens should subsequently be avoided. Inotuzumab may also be preferred in patients who are intended to proceed to subsequent CAR-T therapy using an anti-CD19 construct (see below), in order to minimize the risk of preselection of a CD19 negative subclone. However, conventional chemotherapy or blinatumomab may be preferred in patients who are at risk of VOD/SOS (patients who are early post-HSCT or have pre-existing liver issues).

Patients with relapsed BCR-ABL positive B-ALL on imatinib may be reinduced with a second generation TKI such as dasatinib + chemotherapy. Mutational analysis should be performed at relapse and, if a T315I mutation is detected, ponatinib should be used. Those failing a second generation TKI, should be treated with either inotuzumab or blinatumomab, optimally combined with ponatinib.

Patients with relapsed/refractory T-ALL should receive either salvage chemotherapy with a non-cross resistant regimen (e.g. Hyper-CVAD) or nelarabine.

Regardless of salvage therapy, subsequent relapse is inevitable unless the patient proceeds to allogeneic HSCT or CAR T-cell therapy (see below).

CAR T-Cell Therapy
Chimeric antigen receptor (CAR) T-cell therapy is a treatment in which T lymphocytes are removed from a patient via apheresis, transfected ex vivo with a gene rendering them immunogenic against certain cancer cells, grown and subsequently reinfused into the patient. The activated T-cells then circulate, attack and kill the cancer cells. This treatment has demonstrated considerable activity in patients with relapsed and refractory B-ALL.

Most studies to date have utilized anti-CD19 CAR T-cells. Tisagenlecleucel is the first such therapy to be approved in children and young adults up to age 25 with relapsed/refractory B-ALL. Complete remission rates of 81% were reported,\textsuperscript{152} with a one-year event-free survival (EFS) of 50%, in a cohort of multiple relapsed patients, many of whom had previously received allogeneic HSCT.
Another study of 53 multiply relapsed and refractory B-ALL patients, including many older patients age 30-70 years, from Memorial Sloan Kettering Cancer Center (MSKCC) using a similar anti-CD19 CAR T-cell product, demonstrated an 83% CR rate, a median EFS of 6.1 months and OS of 12.9 months (Park et al, 2021). In this study, patients with a low disease burden (defined as <5% BM blasts) at the time of CAR T infusion had a superior EFS and OS as compared with those who had >5% BM blasts at time of infusion. The ZUMA-3 study, using a different anti-CD19 CAR T-cell product, reported a CR rate of 83% and a median remission duration of 17.6 months (Shah et al, 2021).

CAR T-cell therapy is associated with significant early toxicities, including cytokine release syndrome (CRS), sometimes requiring ICU support +/- tocilizumab, and neurotoxicity; patients therefore require close monitoring in the first two weeks. The MSKCC group and others have reported that the severity of these toxicities is greater in patients with higher tumour burden at the time of CAR-T infusion.

CAR T-cell therapy has become available in Alberta as of 2021, in both Edmonton and Calgary, using both commercial and local investigational products. It is currently being used for patients who have failed at least two different treatment regimens; however, the field is rapidly evolving, and it is likely that it will be evaluated at earlier stages of disease. As some patients relapse with CD19-negative disease, other studies are investigating the use of dual anti-CD19/CD22 CAR-T products.

Potential candidates for this treatment should be referred early for apheresis, prior to administering salvage immunosuppressive chemotherapy, so as not to negatively interfere with the quality of the apheresis product. Most patients will the need some bridging cytoreductive therapy. The latter has a dual purpose: (1) to prevent clinical deterioration while the CAR T product is being prepared, and (2) to reduce the disease burden, potentially reducing the toxicity of the procedure and the risk of subsequent relapse. Optimal cytoreductive strategies are unclear; depending on the patient, it may include non-intensive chemotherapy, inotuzumab or blinatumomab. The latter, although less toxic than other treatments, could potentially increase the risk of emergence of a CD19 negative subclone, so most experts recommend avoiding this if possible. Inotuzumab should be avoided in patients at higher risk of VOD/SOS, including those who are early post-HSCT or who have pre-existing liver disease.

**Recommendations for CAR-T:**
CAR T-cell therapy is indicated for fit patients with B-ALL who have relapsed after allogeneic HSCT, are not otherwise candidates for HSCT, or who have refractory disease. Early referral to the centre’s Cellular Therapy Program is recommended. Patient should not receive salvage T-cell depleting immunosuppressive therapies such as corticosteroids or cyclophosphamide until after apheresis, so as not to impair the quality of the collection. Following apheresis, bridging/cytoreductive therapy should be given, particularly for high tumour burden or rapidly progressive disease, with an aim to
control disease-related complications and reduce the overall tumour burden, while avoiding excessive treatment-related toxicities.

**Future directions**

The treatment landscape of ALL has undergone major changes in the past 5 years, and it is likely that further changes will occur as new information becomes available.

A number of multicenter randomized clinical trials have been evaluating the role of blinatumomab and inotuzumab, in combination with standard frontline chemotherapy regimens, in both younger and older patients with B-ALL. Results from these trials should be available in the next 2-3 years and, if positive, may move these agents into frontline treatment protocols.

For BCR-ABL positive ALL, the combination of ponatinib with Hyper-CVAD chemotherapy has shown very favourable results, without transplant, compared with historical cohorts who had received first of second generation TKI’s. More intriguingly, chemo-free protocols, using blinatumomab combined with either dasatinib or ponatinib, have shown very encouraging results in early studies. If successful, these may fundamentally change how these patients are treated in the future, and may render transplants unnecessary.

As described above, CAR-T cell therapy has shown considerable activity in multiply relapsed and refractory B-ALL. Dual CAR-T constructs, targeting both CD19 and CD22, are in clinical trials, and offer the promise of lowering relapse rates. Moreover, CAR-T therapies will be evaluated in earlier stages of the disease, after first relapse and, eventually, in first CR for high-risk patients with MRD positive disease. This may, if successful, potentially replace the need for allogeneic HSCT for many patients, thereby eliminating the problems associated with GVHD.
References


## Appendix A: Original DFCI Protocol 91-01 (Used for pediatric patients)

<table>
<thead>
<tr>
<th>Phase of Therapy</th>
<th>Time Period</th>
<th>Chemotherapy</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Induction</strong></td>
<td>28 Days</td>
<td>Vincristine 1.5 mg/m2/dose IV, maximum 2 mg, days 3, 10, 17, 24; prednisone 40 mg/m2/d IV/PO for 28 days; doxorubicin 30 g/m2/dose IV, days 1 and 2; methotrexate 4 g/m2 IV for one dose on day 3 IT cytarabine, dosed by age, one dose on day 0 IT cytarabine, dosed by age, one dose on day 17</td>
</tr>
<tr>
<td><strong>CNS Therapy</strong></td>
<td>3 Weeks</td>
<td>SR girls: IT methotrexate/cytarabine for 4 doses during 2 weeks, then every 18 weeks SR boys and all HR patients: cranial XRT 18 Gy, randomly assigned to hyperfractionated (0.9 Gy bid) or conventional (1.8 Gy daily) with IT methotrexate/cytarabine for 4 doses during 2 weeks</td>
</tr>
<tr>
<td><strong>Intensification</strong></td>
<td>Every 3 weeks for 30 weeks</td>
<td>SR: vincristine (2 mg/m2 IV every 3 weeks, maximum 2 mg); dexamethasone 6 mg/m2/d PO for 5 days; methotrexate 30 mg/m2 IV or IM every week; mercaptopurine, randomly assigned to high-dose 1,000 mg/m2 IV for 20 hours, weeks 1 and 2) or conventional 50 mg/m2/d PO for 14 days Asparaginase, randomly assigned to PEG 2,500 IU/m2 IM every 2 weeks for 15 doses or E. coli 25,000 IU/m2 IM every week for 30 doses HR: same as SR patients except dexamethasone 18 mg/m2/d PO for 5 days; no methotrexate; doxorubicin 30 mg/m2 IV every 3 weeks, to total cumulative dose 360 mg/m2, randomly assigned to continuous infusion during 48 hours versus IV bolus</td>
</tr>
<tr>
<td><strong>Continuation</strong></td>
<td>Every 3 weeks until 2 years of continuous complete remission</td>
<td>SR: vincristine 2 mg/m2 IV every 3 weeks, maximum 2 mg; dexamethasone 6 mg/ m2/d PO for 5 days; methotrexate 30 mg/ m2 IV or IM every week; mercaptopurine 50 mg/m2/d PO for 14 days HR: same as SR patients except dexamethasone 18 mg/m2/d PO for 5 days</td>
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## Appendix B: Canadian NCIC DFCI AL.4 Trial

<table>
<thead>
<tr>
<th>Phase of Therapy</th>
<th>Time Period</th>
<th>Chemotherapy</th>
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<tbody>
<tr>
<td><strong>Induction</strong></td>
<td>28 Days</td>
<td>Vincristine 2 mg weekly, days 1, 8, 15 and 22&lt;br&gt;Prednisone 40 mg/m2/day, days 1 –28&lt;br&gt;Doxorubicin 30 mg/m2/dose, days 1 and 2&lt;br&gt;Methotrexate 4 g/m2 (8–24 h after doxorubicin) with leucovorin rescue on day 3&lt;br&gt;E coli L-asparaginase 25 000 IU/m2 IM × 1 dose, day 5&lt;br&gt;IT cytarabine 50 mg, day 0 (prior to initiation of systemic therapy)&lt;br&gt;IT methotrexate (12 mg)/cytarabine (40 mg)/hydrocortisone (50mg), days 15 and 29</td>
</tr>
<tr>
<td><strong>CNS Therapy</strong></td>
<td>3 Weeks</td>
<td>Vincristine 2 mg × 1 dose&lt;br&gt;6-mercaptopurine (6-MP) 50 mg/m2/day orally, × 14 consecutive days&lt;br&gt;Doxorubicin 30 mg/m2 × 1 dose&lt;br&gt;IT methotrexate/cytarabine twice weekly × 4 doses&lt;br&gt;Cranial radiation</td>
</tr>
<tr>
<td><strong>Intensification</strong></td>
<td>Every 3 weeks for 30 weeks</td>
<td>Every 3-week cycles:&lt;br&gt;Vincristine 2 mg, day 1&lt;br&gt;Dexamethasone 18 mg/m2/day b.i.d., orally, days 1 –5&lt;br&gt;Doxorubicin 30 mg/m2, day 1 of each cycle to a (cumulative dose 300 mg/m2)&lt;br&gt;6-MP 50 mg/m2/day orally × 14 consecutive days&lt;br&gt;E. coli asparaginase&lt;br&gt;Individualized dosing: 12 500 IU/m2/dose (starting dose)&lt;br&gt;Methotrexate 30 mg/m2 i.v. or IM weekly, 1 day after asparaginase (no weekly methotrexate until doxorubicin completed&lt;br&gt;IT methotrexate/cytarabine/hydrocortisone at start of a cycle&lt;br&gt;IT therapy consisting of methotrexate/cytarabine at start of a cycle every 18 weeks</td>
</tr>
<tr>
<td><strong>Continuation</strong></td>
<td>74 weeks</td>
<td>Same as intensification except no asparaginase and dexamethasone dose reduced to 6 mg/m2/day</td>
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**Appendix C: Princess Margaret Hospital Modified DFCI 91-01/Al.4 Protocol** for BCR-ABL1 negative adult patients aged <60 Years, Modified for Pegaspargase (Native *E. coli* asparaginase no longer available)

<table>
<thead>
<tr>
<th>Phase of Therapy</th>
<th>Time Period</th>
<th>Chemotherapy</th>
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</table>
| **Induction**    | 28 Days     | Vincristine 2 mg weekly, days 1, 8, 15 and 22  
Prednisone 40 mg/m2/day, days 1–28  
Doxorubicin 30 mg/m2/dose, days 1 and 2  
Methotrexate 4 g/m2 (8–24 h after doxorubicin) with leucovorin rescue on day 3  
Pegasparagase 2000 U/m2 IV × 1 dose, day 5  
IT therapy consisting of methotrexate (12 mg)/cytarabine (50 mg)/hydrocortisone (15 mg), days 1, 15, 29 |
| **CNS Therapy**  | 3 Weeks     | Vincristine 2 mg × 1 dose day 1  
6-mercaptopurine (6-MP) 50 mg/m2/day orally, day 1–14  
Doxorubicin 30 mg/m2 × 1 dose day 1  
IT therapy consisting of methotrexate (12 mg)/cytarabine (50 mg)/hydrocortisone (15 mg) twice weekly × 4 doses |
| **Intensification** | Every 3 weeks for 30 weeks | Vincristine 2 mg, day 1  
Dexamethasone 9 mg/m2 PO BID, days 1–5  
Doxorubicin 30 mg/m2, day 1 of cycles 1–7 (to a cumulative dose 300 mg/m2)  
6-MP 50 mg/m2/day orally × 14 consecutive days  
Pegasparagase 2000 IU/m2 IV day 1  
Methotrexate 30 mg/m2 IV, IM or PO weekly on day 2, 9, and 16 on cycles 8–10 (after maximum doxorubicin dose completed).  
IT therapy consisting of methotrexate (12 mg)/cytarabine (50 mg)/hydrocortisone (15 mg) start of a cycle every 18 weeks |
| **Continuation** | 74 weeks    | Vincristine 2 mg, day 1  
Dexamethasone dose reduced to 6 mg/m2 BID  
6-MP 50 mg/m2/day orally × 14 consecutive days  
Methotrexate 30 mg/m2 IV, IM or PO weekly days 1, 8 and 15  
IT therapy consisting of methotrexate/cytarabine at start of a cycle every 18 weeks |
Appendix D: Princess Margaret Hospital Modified DFCI 91-01/Al.4 Protocol\textsuperscript{20} for BCR-ABL1 negative adult patients aged >60 Years, Modified for Pegaspargase (Native \textit{E. coli} asparaginase no longer available)

<table>
<thead>
<tr>
<th>Phase of Therapy</th>
<th>Time Period</th>
<th>Chemotherapy</th>
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</thead>
</table>
| **Induction**    | 28 Days           | Vincristine 2 mg weekly, days 1, 8, 15  
Dexamethasone 40 mg, days 1-4, 9–12  
Doxorubicin 30 mg/m2/dose, days 1and 2  
Methotrexate 40 mg/m^2, day 3  
Pegasparagase 1000 U/m2 IV × 1 dose, day 4  
IT Cytarabine 70 mg, day 1  
IT therapy consisting of methotrexate (12mg)/cytarabine (50 mg)/hydrocortisone (15 mg), days 15, 29 |
| **CNS Therapy**  | 3 Weeks           | Vincristine 2 mg × 1 dose day 1  
6-mercaptopurine (6-MP) 50 mg/m2/day orally, × 14 consecutive days (day 1 -14)  
Doxorubicin 30 mg/m2 × 1 dose day 1  
IT therapy consisting of methotrexate (12mg)/cytarabine (50 mg)/hydrocortisone (15 mg) twice weekly × 4 doses |
| **Intensification** | Every 3 weeks for 21 weeks | Vincristine 2 mg, day 1  
Dexamethasone 6 mg b.i.d., orally, days 1–5  
Doxorubicin 30 mg/m2, day 1  
6-MP 50 mg/m2/day orally × 14 consecutive days  
Pegasparagase 1000 U/m2 IV day 1  
IT therapy consisting of methotrexate (12mg)/cytarabine (50 mg)/hydrocortisone (15 mg) start of a cycle every 18 weeks |
| **Continuation** | 74 weeks          | Vincristine 2 mg, day 1  
Dexamethasone 6 mg PO BID days 1-5  
6-MP 50 mg/m2/day orally × 14 consecutive days  
Methotrexate 30 mg/m2 IV, IM or PO weekly days 1, 8 and 15  
IT therapy consisting of methotrexate/cytarabine at start of a cycle every 18 weeks |
### Appendix E: Princess Margaret Hospital Modified DFCI 91-01/AL.4 Protocol for Philadelphia Chromosome/BCR-ABL1 Positive Adult Patients Aged <60 Years

<table>
<thead>
<tr>
<th>Phase of Therapy</th>
<th>Time Period</th>
<th>Chemotherapy</th>
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</table>
| **Induction**    | 28 Days     | Vincristine 2 mg, weekly, days 1, 8  
|                  |             | Vinblastine 10 mg, day 15  
|                  |             | Prednisone 40 mg/m2/day, days 1–28  
|                  |             | Doxorubicin 30 mg/m2/dose, days 1 and 2  
|                  |             | Methotrexate 4 g/m2 (8–24 h after doxorubicin) with leucovorin rescue on day 3  
|                  |             | IT cytarabine 70 mg, day 1  
|                  |             | IT therapy consisting of methotrexate (12 mg)/cytarabine (50 mg)/hydrocortisone (15 mg), days 15, 28  
|                  |             | Imatinib 400 mg, days 1-16  |
| **CNS Therapy**  | 3 Weeks     | Vinblastine 10 mg × 1 dose, day 1  
|                  |             | 6-mercaptopurine (6-MP) 50 mg/m2/day orally, × 14 consecutive days (day 1-14)  
|                  |             | Doxorubicin 30 mg/m2 × 1 dose, day 1  
|                  |             | IT therapy consisting of methotrexate (12 mg)/cytarabine (50 mg)/hydrocortisone (15 mg) twice weekly × 4 doses  
|                  |             | Imatinib 400 mg daily, days 1-21  |
| **Intensification** | Every 3 weeks for 21 weeks | Vinblastine 10 mg, day 1  
|                  |             | Dexamethasone 9 mg/m2/ b.i.d., orally, days 1–5  
|                  |             | Doxorubicin 30 mg/m2, day 1 of each cycle to a cumulative dose 300 mg/m2  
|                  |             | 6-MP 50 mg/m2/day orally × 14 consecutive days  
|                  |             | Methotrexate 30 mg/m2 i.v. or IM weekly, days 1, 8 and 15 (after maximum doxorubicin dose completed)  
|                  |             | IT therapy consisting of methotrexate (12 mg)/cytarabine (50 mg)/hydrocortisone (15 mg) every 18 weeks  
|                  |             | Imatinib 400 mg, days 1-21  |
| **Continuation** | 74 weeks    | Vinblastine 10 mg, day 1  
|                  |             | Dexamethasone 6 mg/m2 BID, days 1-5  
|                  |             | 6-MP 50 mg/m2/day orally × 14 consecutive days  
|                  |             | Methotrexate 30 mg/m2 i.v. or IM weekly days 1, 8 and 15  
|                  |             | IT therapy consisting of methotrexate (12 mg)/cytarabine (50 mg)/hydrocortisone (15 mg)  
|                  |             | at start of a cycle every 18 weeks  
|                  |             | Imatinib 400 mg, days 1-21  |
| **Long Term Maintenance** |          | Imatinib 600 mg daily |
### Appendix F: GRAAPH-2005 Induction Protocol for Philadelphia Chromosome/ BCR-ABL1 Positive Adult Patients,\textsuperscript{132} with Modification for Older Patients

<table>
<thead>
<tr>
<th>Phase of Therapy</th>
<th>Time Period</th>
<th>Chemotherapy</th>
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</table>
| Prephase         | 7 Days      | Prednisone 60 mg/m2/day, days -7 to -1  
IT therapy consisting of methotrexate (12mg)/cytarabine (50 mg)/hydrocortisone (15 mg) |
| Cycle 1          | 4 Weeks     | Vincristine 2 mg IV, days 1, 8, 15, 22  
(for pts age $\geq$ 65, days 1 and 15 only)  
Dexamethasone 40 mg/d PO, days 1-2, 8-9, 15-16, 22-23  
(for pts age $\geq$ 65, 20 mg/d)  
Imatinib 400 mg daily, days 1-28  
IT therapy consisting of methotrexate (12mg)/cytarabine (50 mg)/hydrocortisone (15 mg), days 1, 8, 15  
Filgrastim 5 μg/kg/d sc daily, days 15 until neutrophil recovery |
| Cycle 2 (only for pts < age 65) | 3 Weeks | Methotrexate 1000 mg/m2/d CIV over 24 hours, day 1  
Cytarabine 3000 mg/m2/d12h IV x 4 doses, days 2-3  
(1500 mg/m2 for pts age 60-64)  
Imatinib 400 mg PO BID, days 1-14  
Filgrastim 5 μg/kg/d sc daily, days 9 until neutrophil recovery |

**Should be followed by Intensification and maintenance therapy** (e.g. DFCI as per Appendix E or HyperCVAD + Imatinib)
Development and Revision History
This guideline was developed by a multidisciplinary working group comprised of members from the Alberta Provincial Hematology Tumour Team, external participants identified by the Working Group Lead, and a methodologist from the Guideline Resource Unit. The draft guideline was externally reviewed and endorsed by members of the Alberta Provincial Hematology Tumour Team who were not involved in the guideline’s development, including surgical oncologists, radiation oncologists, medical oncologists, hematologists, nurses, pathologists, and pharmacists. A detailed description of the methodology followed during the guideline development process can be found in the Guideline Resource Unit Handbook.

This guideline was originally developed in 2016.

Maintenance
A formal review of the guideline will be conducted in 2023. If critical new evidence is brought forward before that time, however, the guideline working group members will revise and update the document accordingly.

Abbreviations
AHS, Alberta Health Services; CCA, Cancer Care Alberta

Disclaimer
The recommendations contained in this guideline are a consensus of the Alberta Provincial Hematology Tumour Team and are a synthesis of currently accepted approaches to management, derived from a review of relevant scientific literature. Clinicians applying these guidelines should, in consultation with the patient, use independent medical judgment in the context of individual clinical circumstances to direct care.

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Funding Source
Financial support for the development of Cancer Care Alberta’s evidence-based clinical practice guidelines and supporting materials comes from the Cancer Care Alberta operating budget; no outside commercial funding was received to support the development of this document.

All cancer drugs described in the guidelines are funded in accordance with the Outpatient Cancer Drug Benefit Program, at no charge, to eligible residents of Alberta, unless otherwise explicitly stated. For a complete list of funded drugs, specific indications, and approved prescribers, please refer to the Outpatient Cancer Drug Benefit Program Master List.

Conflict of Interest Statements
Dr. Joseph Brandwein reports advisory board participation for Pfizer, Abbvie, Taiho Pharma, Jazz Pharma, BMS/Celgene, Avir Pharma, Astellas, and Amgen.

Derek Tilley has nothing to disclose.

Citation