Scanning Label or Accession # (lab only)

	BERTA P			OLEC	ULAK P	ATHOL	.UGY R	EQUIS	SITION							
	LABORA ders in Labor		ine													
PHN Date of Birth									<i>-уууу)</i>							
	Expiry:															
eni	Legal Las	Lega	egal First Name				IV	Middle Name								
Patient	Alternate Identifier Prefer				_			Male Non-bii	nary 🔲	=	Female Prefer not to disclos		Phone se			
	Address	City/	City/Town			Prov				Postal Code						
(s	Authorizing Provider Name (last, first, middle)							Сор	Copy to Name (last, first, middl			dle)	Copy to Name (last, first, middle)			
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Provider (s	CC Provider ID CC Submitter II			D Lega	D Legacy ID			ne		F			Phone			
P	Clinic Name						Clini	c Name	Clinic Na			ic Name				
Col	lection	ection Date (dd-Mon-yyyy)			Time (24 hr)			Location			Co			Collector ID		
*H&E(s) included?  *% viable neopla								? Yes No Consult Block (s) No.:								
*Blood/BM													Tissue/Fluid Source:			
	d/BM specimen		bmitted in El	DTA or AC	D tubes.	**Liquid Bio	opsy in Sti	eck tubes	ONLY.							
	ory/Diagnosis			<b> </b>				0	C	:- A	lta nati a na					
Lymphocyte Receptor Gene Rearrangement/Translocation  B-cell Receptor IGH Gene Rearrangement									Cancer Genomic Alterations							
	B-cell Receptor IGH Gene Rearrangement T-Cell Receptor								Mutation Analysis  ☐ EGFR, BRAF, KRAS and PIK3CA (Lung Panel)							
	Bol-1 t(11;14)								T790M EGFR (cfDNA - Blood only)							
	Bcl-2 t(14;18)								☐ KRAS, NRAS, BRAF, EGFR and PIK3CA (Colon Panel)							
	gH Somatic H		☐ BRAF, NRAS, KRAS, HRAS, KIT, GNA11, GNAQ, PTEN, RAC,							RAC,						
	and Bacteria					RPS27 and TERT (Melanoma Panel)										
	Cytomegalovi	rus						☐ IDH1, IDH2, H3.3, TERT, BRAF, PTEN, TP53 (CNS/Neuro Panel)								
□ +	Herpes Simplex Virus (HSV 1 & 2)								☐ CTNNB1, GNAS, KIT, PDGFRA, IDH1 and IDH2 (Sarcoma Pane							
	aricella Zoster Virus								☐ ThyroSPEC ™ (Thyroid Cytology Panel)							
	pstein-Barr Virus								☐ BRCA1 / BRCA2 Somatic mutations (Gyneoncology)							
□ F	arvovirus B19								Translocation Analysis							
	Mycobacteriun			ALK, ROS and RET (Lung Fusion Panel, Only for confirmation of IHC)												
□ +	Human Polyo			Microsatellite Analysis												
Human Papillomavirus (Low risk 6,11,42,81: High risk 16, 18, 31, 33, 34, 35, 39, 45, 51, 52, 53, 56, 58,59, 66, 67, 68, 70, 73, 82)									<u> </u>							
									Epigenetics (Methylation)							
									☐ MGMT Promoter Methylation							
								Hum	Human DNA Identity Testing							
Received: In <u>Comments (lab use only)</u>							!		Mol Path#							

SEE REVERSE (PAGE 2) FOR SAMPLE REQUIREMENTS, INSTRUCTIONS ON SUBMITTING A SPECIMEN, AND SHIPPING ADDRESS. FAX REQUISITION TO THE MOLECULAR PATHOLOGY ACCESSIONING DESK AT 403-944-4748

\_ In. \_

Sent:

## **Alberta Precision Laboratories**

# www.albertaprecisionlabs.ca

Main Reception 403-770-3500 Laboratory Information Centre 403-770-3600

Medical Staff: For test information, specimen collection instructions, etc. see <a href="https://www.albertaprecisionlabs.ca">www.albertaprecisionlabs.ca</a>
Physicians may contact the Laboratory Information Centre (L.I.C.) 403-770-3600 for test results and related inquiries

FOR GENERAL CORRESPONDENCE: EMAIL fmc.molpathtechs@albertaprecisionlabs.ca

## **Specimen Requirements**

Microsatellite Instability (MSI) and Loss of Heterozygosity (LOH) testing require the suspected cancer tissue to be paired with normal tissue (preferably from an independent block with no tumour present) or blood from the patient (blood must be sent in EDTA or Sodium Citrate ACD vacutainers, 2mL minimum). MSI analysis may be performed if no normal is available (LOH ANALYSIS WILL NOT BE PERFORMED WITHOUT NORMAL TISSUE FOR COMPARISON.)

**Cancer Genomic Alteration** testing requires an H&E stained slide, with the tumour rich area clearly indicated, sent with the block for testing in order to facilitate microdissection of the sample if required. An assessment of tumour cellularity is required. Enter the percentage (%) of malignant cells in the indicated region or for the entire specimen. Refer to page 1 of the requisition. To avoid delays in result reporting, the H&E slide should be representative of the block being submitted for testing.

**T790M EGFR mutation test** is performed on blood samples only. Samples must be collected at the Special Service Building (SSB) Outpatient Laboratory of the Foothills Hospital using Streck cfDNA blood collection tubes. Note that this test **will not be performed on FFPE samples. All such requests will default to Lung Panel testing**.

**Human DNA Identity Testing** for specimen misidentification, floater, contamination identification, etc. requires comparison specimens to evaluate genetic identity. Indicate on the requisition if the submitted block is the reference or the unknown block/specimen. If submitting blocks from two patients as possible sources of contaminant, each patient requires a separate or different requisition. Indicate on both requisitions the name of the other patient involved in the test.

**Lung Fusion Panel** testing will **only be performed to confirm positive IHC result**. Indicate under History/Diagnosis that IHC has been performed and was positive for either ALK, RET or ROS.

### TEST REQUESTS FROM NON-APL LABORATORIES

#### Frozen, fresh, or other perishable specimens:

Please contact the Molecular Pathology Accessioning Desk directly by phone at 403-944-4542, fax at 403-944-4748 or email <a href="mailto:fmc.molpathtechs@albertaprecisionlabs.ca">fmc.molpathtechs@albertaprecisionlabs.ca</a> to arrange delivery of specimen.

### Fixed (paraffin-embedded or fixed cytology):

1. Specimens must be sent to the Consult Desk, Alberta Precision Laboratories

c/o Consult Desk, APL AP/Cytology 7<sup>th</sup> floor, McCaig Tower 1403 - 29<sup>th</sup> Street NW Calgary, AB T2N 2T9

Phone: 403-944-4542 Fax: 403-944-4748

- All specimens must be accompanied by a fully completed Molecular Pathology requisition.
- Please note specimen requirements for the requested test (see above).
- 4. Note: All inquiries for test results should be directed to the Laboratory Information Centre (L.I.C.) at 403-770-3600.

**DO NOT** send specimens or requests to the Laboratory Director, Dr. A. Box

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